

**Supplementary materials:**

**Revealing the neurobiology underlying interpersonal neural synchronization  
with multimodal data fusion**

Leon D. Lotter, Simon H. Kohl, Christian Gerloff, Laura Bell, Alexandra Niephaus,  
Jana A. Kruppa, Juergen Dukart, Martin Schulte-Rüther, Vanessa Reindl, Kerstin Konrad

Published in *Neuroscience & Biobehavioral Reviews*  
(<https://doi.org/10.1016/j.neubiorev.2023.105042>)

**Table of Contents**

<b>1</b>	<b>Supplementary Materials and Methods.....</b>	<b>3</b>
1.1	Literature search and data extraction .....	3
1.1.1	Search string.....	3
1.1.2	Study inclusion criteria .....	3
1.1.3	Extracted study information.....	4
1.1.4	Sample data extraction.....	4
1.1.5	fMRI coordinate data extraction .....	5
1.1.6	fNIRS coordinate data extraction and reconstruction.....	5
1.2	Calculation of the fail-safe-N.....	6
1.3	Modelling of spatial uncertainty in fNIRS meta-analyses .....	6
1.4	Resting-state fMRI data processing .....	7
1.5	Relative and absolute distributions within major resting-state networks.....	7
1.6	Processing of Allen Brain Atlas mRNA expression data.....	8
1.7	Validation analyses for in-vivo INS-GABA <sub>A</sub> associations.....	8
1.8	Dominance analysis on nuclear imaging and neuronal cell type data.....	9
1.9	Clustering and visualization of INS-associated GeneOntology categories.....	9
<b>2</b>	<b>Supplementary Results .....</b>	<b>10</b>
2.1	FMRI meta-analysis sensitivity analyses .....	10
2.1.1	Influence of individual experiments .....	10
2.1.2	Risk of publication bias .....	10
2.2	FNIRS meta-analysis sensitivity analyses.....	10
2.2.1	Alternative “fNIRS-Indices”.....	10
2.2.2	Modelling of spatial uncertainty .....	11
2.2.3	Restricted set of fNIRS experiments.....	11
2.3	MACM sensitivity analyses .....	11
2.3.1	Controlling for baseline activation probability .....	11
2.3.2	Spatial alignment between INS-ALE and MACM activation patterns.....	12
2.4	Spatial relationships to established resting-state networks .....	12
2.5	Neurotransmitter-associations sensitivity analyses.....	12
2.5.1	GABA <sub>A</sub> -related mRNA expression .....	12
<b>3</b>	<b>Supplementary References .....</b>	<b>13</b>
<b>4</b>	<b>Supplementary Tables .....</b>	<b>19</b>
	Table S1: fMRI experiments included for meta-analysis.....	19
	Table S2: fNIRS experiments included for meta-analysis .....	22
	Table S3: Sources of neurotransmitter maps .....	23

Table S4: Gene category enrichment datasets .....	24
Table S5: Information on clusters resulting from ALE and MACM analyses .....	25
Table S6: Region-wise fNIRS meta-analyses.....	28
Table S7: Functional-decoding results .....	30
Table S8: Results of neurotransmitter association analyses .....	31
Table S9: Results of gene category enrichment analyses.....	34
Table S10: Results of GeneOntology category clustering.....	35
<b>5 Supplementary Figures.....</b>	<b>36</b>
Figure S1: INS foci reported in each fMRI experiment .....	36
Figure S2: INS foci reported in each fNIRS experiment.....	38
Figure S3: ALE and MACM sensitivity analyses .....	39
Figure S4: Spatial overlap with prior meta-analytically derived networks and rTPJ subunits .....	40
Figure S5: Complete meta-analytic results of INS fNIRS experiments.....	41
Figure S6: Spatial associations of INS to neurotransmitter distributions.....	42
Figure S7: Validation of INS-GABA <sub>A</sub> associations using mRNA expression data .....	43
Figure S8: Variance of the INS distribution explained by significantly associated nuclear imaging and neuronal cell type data.....	44
Figure S9: Clustering of GeneOntology biological process categories spatially associated with INS .....	45

## 1 Supplementary Materials and Methods

### 1.1 Literature search and data extraction

#### 1.1.1 Search string

```
(([hyperscanning] OR
[neural entrainment] OR
([brain-to-brain] OR
[interbrain] OR
[inter-brain] OR q
[interpersonal])) AND
([synchrony] OR
[synchronization] OR
[coupling] OR
[connectivity] OR
[networks] OR
[alignment] OR
[coherence])) AND
([fNIRS] OR
[NIRS] OR
[near-infrared spectroscopy] OR
[fMRI] OR
[MRI] OR
[magnetic resonance imaging]))
```

#### 1.1.2 Study inclusion criteria

- **Article type:** original research, peer-reviewed or preprint, English language (exclude: conference paper, review, meta-analysis, opinion)
- **Subjects:** human, healthy, between 18 and 65 years
- **Imaging Technology:** fNIRS or fMRI
- **Methodological study design:** “hyperscanning” or “pseudohyperscanning”
- **Task:** uni- or bidirectional interaction between  $\geq$  two subjects
- **Analysis:** temporal synchrony between data derived from hemodynamic signals of  $\geq$  two subjects
  - **fMRI:** analysis of data on whole-brain level (exclude: “region of interest” approaches)
  - **fNIRS:** analysis of channel-wise signals (exclude: data analysis after averaging data from multiple channels to form a “region of interest”)
- **Data availability:**
  - **fMRI:** reporting of coordinates in Montreal Neurological Institute (MNI) or Talairach space, or availability of coordinates or 3D volumes on request
  - **fNIRS:** reporting of coordinates in MNI or Talairach space, or availability of coordinates or fNIRS probe setups on request, or reporting of  $\geq$  one coordinate in

international 10-10 system, alignment to a reference plane, and usage of a square probe setup, thus allowing for reconstruction of the probe positioning

### ***1.1.3 Extracted study information***

- **General:** authors, publication year, title, journal, identifier
- **Sample:** number of subjects, number of dyads, male/female ratio, relationship between interacting subjects, mean age, age standard deviation, age range
- **Method:** imaging technology, hyperscanning or pseudohyperscanning, general analysis technique, applied contrasts
  - **fMRI:** repetition time, time lag, analysis frequency band, analysis level (voxel-wise, independent component analysis, seed-to-voxel, whole-brain parcellation), analysis software
  - **fNIRS:** device, wavelength of emitted light, sampling rate, analyzed data (HbO, HbR), analysis frequency band, time lag, optode array format and positioning
- **Task and setting:** general domain, exact applied task, setting
  - **fMRI:** (live) audio/video contact
  - **fNIRS:** positioning of subjects to each other (side-by-side, face-to-face, separated)
- **Coordinate data:** coordinates in MNI or Talairach space, author contact information if no coordinates were available
  - **fNIRS:** source of coordinates (virtual registration, digitization, MRI-registration)

### ***1.1.4 Sample data extraction***

Sample data were extracted as reported in included studies. If a study reported exclusion of subjects, e.g., due to low data quality, sample sizes after exclusion of these subjects were retained. When, e.g., in case of pseudo-hyperscanning fMRI and some fNIRS studies, demographic data was provided for the study sample and a small group of “special” participants who were scanned repeatedly, sample sizes were determined by the overall sum of participants, while the number of dyads equaled the number of formed dyads, and age mean and standard deviation were calculated as weighted averages. For example, if a study included  $n = 20$  students (mean age 20 years) and  $n = 2$  teachers (mean age 24 years), forming  $n = 20$  dyads with teachers being scanned repeatedly, we set the sample size to 22, the number of dyads to 20, and the mean age to 20.37 years.

When a study split its sample in multiple groups with independent subjects and independently analyzed data, these groups were included as separate experiments. When sample information (i.e., male/female ratio, age) were reported only for the whole sample but not for subgroups, this information was applied to all subgroups at the risk of small inaccuracies (which only apply to the results Tables S1 and S2 but have no influence on our meta-analytic results).

When samples between different studies overlapped, i.e., data derived from the same subjects was analyzed and published multiple times, these data, and reported brain coordinates, were concatenated into one experiment. If it was most likely that (according to the descriptions of sample characteristics, tasks, and the publishing authors), a sample was analyzed twice but this was not mentioned in the papers and the authors did not answer requests, we chose a conservative approach

and concatenated respective data into one experiment. When concatenating data, the lowest reported sample size of all individual studies was used for the resulting “experiment” to be included in meta-analyses.

### ***1.1.5 fMRI coordinate data extraction***

Coordinates provided in MNI space were not modified. Coordinates in Talairach space were converted to MNI using the Lancaster transform (1).

When authors sent thresholded volumetric imaging data, we extracted a maximum of three peak coordinates from each cluster (minimum distance of 10 mm, minimum size of 10 voxels).

When authors sent independent component analysis results in the form of volumetric data depicting independent components, we Z-normalized these maps while excluding zero-voxels, applied a cluster threshold of  $Z > 2.5$  and retained only clusters exceeding a volume of 0.1% of all non-zero in-brain voxels. From all studies applying independent component analysis, for each reported or estimated cluster, a maximum number of three peak coordinates with a minimum distance of 10 mm was retained for meta-analysis.

When data was analyzed using a whole-brain parcellation, for each parcel showing INS, the center of mass was used as focus coordinates.

When authors analyzed INS not only for homologous brain areas (voxels, parcels, independent components) but in a pairwise fashion for the whole brain, foci representing all connected regions were retained for meta-analysis (e.g., when significant INS was found between the right temporoparietal junction of subject 1 and the right insula of subject 2, we would include foci representing both regions).

### ***1.1.6 fNIRS coordinate data extraction and reconstruction***

For studies that did not report coordinates, we obtained MNI coordinates according to the following workflow:

When studies reported to have used the virtual registration method (2) and used a common static rectangular optode placeholder, we used coordinates provided by Tsuzuki et al. ([http://brain-lab.jp/wp/?page\\_id=58](http://brain-lab.jp/wp/?page_id=58)). If studies did not report to have used the virtual registration method but their description of optode placements matched one of the probe setups for which virtual registration results were available, we used these for the analyses [e.g., Cui et al., 2012 (3) matched Fpz\_low3\_HorSag\_3x5].

In some cases, a probe set configuration reported by study A was used in study B as well and study B was published by authors from the same research group or University. When coordinates for study B were available, we used these for study A as well [e.g., for Liu et al, 2019 (4), we used the temporoparietal coordinates from Lu et al, 2020b (5)].

When this was not possible, we reconstructed the probe sets based on the spatial information provided by the studies using AtlasViewer (<https://openfnirs.org/software/homer>) and registered it to a scaled version of the Colin27 MNI atlas. Since head size information were not provided by the studies in question, we could not register the Colin27 atlas in AtlasViewer to study-specific head sizes but had to rely on a standard head size for all reconstructions. In order to get an appropriate approximation of the average head shape, we calculated the average head sizes for white and Asian

women and men (560.01 mm) based on data provided by Ball (6) (<https://repository.tudelft.nl/islandora/object/uuid:2d038418-8923-4605-92e8-ca3df57ea731>) and Harrison & Robinette (7) (<https://apps.dtic.mil/sti/pdfs/ADA406674.pdf>). To be suited for reconstruction, studies had to report at least two anatomical landmarks of the EEG 10-10 or 10-5 system. We rated the “reconstructability” of studies as *high* if EEG channel positions for every single optode were reported, as *medium* if at least two EEG channel positions for each optode array were given, or as *low* if one EEG channel position and some additional spatial information about the alignment of the probe set, e.g., “15° angle between probe patches and transverse plane” was reported. When only one landmark but no additional information or no information at all was reported, these studies were excluded.

When studies analyzed and reported fNIRS data in voxel space and reported significant voxel-level MNI coordinates, we obtained coordinates according to the workflow lined out above and retained, for each reported significant voxel, coordinates of the three fNIRS channels most closely located to this voxel.

## 1.2 Calculation of the fail-safe-N

The robustness against publication bias of each cluster was estimated as the fail-safe-N (8). For each cluster, noise experiments with sample sizes and foci numbers drawn from the actual included fMRI experiments were generated. Foci coordinates for these noise studies were randomly drawn from all gray matter voxels excluding those in the brain quadrant where the respective cluster’s center of mass was located. Based on preset minimum and maximum numbers of added noise experiments (the number of contributing experiments and the number of noise studies needed to reach a minimum contribution of 10% of studies) (8), we iteratively searched for the minimum number of noise experiments needed to render the cluster insignificant while applying standard voxel-level and cluster-level thresholds ( $p < .001$  and  $p < .05$ ). The resulting number mirrors the hypothetical minimum of negative studies that could have “remained in the file drawer” necessary for the respective cluster to fail significance thresholds.

## 1.3 Modelling of spatial uncertainty in fNIRS meta-analyses

To incorporate spatial uncertainty of fNIRS data in fNIRS meta-analyses, we iteratively recalculated both the parcel-wise fNIRS meta-analysis and the combined ALE after randomization of fNIRS coordinates within a 10 mm radius and a strongly constrained cortical MNI-152 template (1,000 iterations). The radius choice was based on previous data showing a localization error of 18 mm in preregistered model-based fNIRS-fMRI registration (9, 10).

To evaluate the fNIRS-only meta-analytic results, we calculated, for each parcel, the percentage of sub-threshold  $p$  values ( $p < .05$ ) relative to the total number of iterations as well as the median  $p$  value resulting from all iterations.

The combined ALE was repeated with randomized coordinates in the same fashion as the main analysis. However, we adopted the cluster mass threshold estimated from the maximum cluster size distribution of the original fNIRS-fMRI ALE instead of recalculating individual thresholds for each fNIRS coordinate randomization iteration to reduce the required computational power.

We then evaluated the results by calculating the proportion of thresholded ALE maps for which nonzero ALE values were present within the clusters derived from the main combined fMRI-fNIRS ALE.

#### 1.4 Resting-state fMRI data processing

To estimate the functional connectivity pattern present within the task-based coactivation network,  $N = 120$  unrelated subjects from the Human Connectome Project S900 release (11) were randomly selected from age and gender groups (20 females and 20 males from each age group: 22–25, 26–30, and 31–35 years). We used the S900 extensively preprocessed volumetric data (ICA-FIX denoised) thoroughly described in the reference manual (12). Further processing in CONN (<https://web.conn-toolbox.org/>) included: resampling of the data to a 3 mm isotropic MNI-152 template, concatenation of the first two resting-state sessions (resulting in 30 min), linear detrending and bandpass filtering (0.01–0.08 Hz), averaging of voxel-wise timeseries across MACM clusters, and calculation of semipartial correlations between clusters (13). After exclusion of subjects exceeding framewise displacement cutoffs of 2 mm translation or 2 degree rotation ( $n = 5$ ), we calculated two-sided one-sample t-tests against zero on the r-to- $Z$  transformed semipartial correlation coefficients representing each (directed) functional connection. To identify the strongest functional connections, the resulting  $p$  value matrix was thresholded at FWE-corrected  $p < .05$  (Bonferroni) and only positive connections were interpreted to exclude potential spurious negative connections introduced through noise regression (14).

#### 1.5 Relative and absolute distributions within major resting-state networks

To characterize the ALE clusters and the associated MACM networks in respect to their spatial overlap with established brain-wide resting-state networks (15–17), we adopted a frequently used method characterizing the absolute and relative distributions of our target volumes across these networks (18). The *relative distribution* refers to the proportion of activated voxels within a reference network compared to all activated voxels, while the *absolute distribution* is calculated as the proportion of activated voxels compared to all voxels within a reference network. To estimate significance of these spatial associations, we then permuted the input coordinates (1,000 repetitions; fMRI INS coordinates and rTPJ-associated BrainMap coordinates, respectively) within a grey-matter mask (MNI-152, thresholded at  $> .2$ ), performed ALE analyses on these coordinates, and iteratively reassessed the resting-state network overlaps to generate null distributions of the overlap metrics. From these null distributions, we calculated empirical  $p$  values denoting the probability of a false positive finding under the null hypothesis of random spatial localizations of INS/MACM coordinates. The  $p$  values were FDR-corrected for each metric across the combined rTPJ and MACM results. To reduce computational cost, we did not run the full cluster mass permutation procedure as in the main ALE/MACM analyses but extracted the cluster mass thresholds from the primary analyses and used these to perform cluster-level inference on the null data after applying a voxel-level threshold of  $p < .001$ .



## 1.6 Processing of Allen Brain Atlas mRNA expression data

Regional microarray expression data were obtained from 6 post-mortem brains (1 female, age range 24.0–57.0 years, mean age  $42.50 \pm 13.38$  years) provided by the Allen Human Brain Atlas (<https://human.brain-map.org>) (19). Data were processed with the abagen toolbox (version 0.1.3; <https://github.com/rmarkello/abagen>) (20) using a volumetric atlas in MNI space covering 116 cortical and subcortical brain regions (21, 22).

First, microarray probes were reannotated using data provided by Arnatkevičiūtė et al. (23); probes not matched to a valid Entrez ID were discarded. Next, probes were filtered based on their expression intensity relative to background noise (24), such that probes with intensity less than the background in  $\geq 50\%$  of samples across donors were discarded, yielding 31,569 probes. When multiple probes indexed the expression of the same gene, we selected and used the probe with the most consistent pattern of regional variation across donors [i.e., differential stability (25)], calculated with:

$$\Delta_S(p) = \frac{1}{\binom{N}{2}} \sum_{i=1}^{N-1} \sum_{j=i+1}^N \rho[B_i(p), B_j(p)]$$

where  $p$  is Spearman's rank correlation of the expression of a single probe,  $p$ , across regions in two donors  $B_i$  and  $B_j$ , and  $N$  is the total number of donors. Here, regions correspond to the structural designations provided in the ontology from the Allen Human Brain Atlas. The MNI coordinates of tissue samples were updated to those generated via non-linear registration using the Advanced Normalization Tools (<https://github.com/chrisfilo/alleninf>). Samples were assigned to brain regions in the provided atlas if their MNI coordinates were within 2mm of a given parcel. All tissue samples not assigned to a brain region in the provided atlas were discarded. Inter-subject variation was addressed by normalizing tissue sample expression values across genes using a robust sigmoid function (26):

$$x_{norm} = \frac{1}{1 + \exp\left(-\left(\frac{x - \langle x \rangle}{IQR_x}\right)\right)}$$

where  $\langle x \rangle$  is the median and  $IQR_x$  is the normalized interquartile range of the expression of a single tissue sample across genes. Normalized expression values were then rescaled to the unit interval:

$$x_{scaled} = \frac{x_{norm} - \min(x_{norm})}{\max(x_{norm}) - \min(x_{norm})}$$

Gene expression values were then normalized across tissue samples using an identical procedure. Samples assigned to the same brain region were averaged separately for each donor and then across donors, yielding a regional expression matrix with 116 rows, corresponding to brain regions, and 15,633 columns, corresponding to the retained genes.

## 1.7 Validation analyses for in-vivo INS-GABA<sub>A</sub> associations

To validate the spatial association found between INS and the GABA<sub>A</sub> receptor distribution, the following list of GABA-related genes was collected (27, 28) and data were extracted from the

Allen Brain atlas: GABRA1, GABRA2, GABRA3, GABRA4, GABRA5, GABRB1, GABRB2, GABRB3, GABRG1, GABRG2, GABRG3, GABRD, GABRE, GAD1, GAD2, PVALB, SST, VIP, CCK, NPY, CALB1, CALB2, NOS1, RELN, ADRB2, LHX6, TAC1, TAC3, TAC4, SLC6A1, SLC6A13, SLC6A12, SLC32A1, GABBR1, GABBR2.

Following genes were not available after preprocessing of Allen Brain Atlas data: GABRA6, GABRP, GABRQ, GABRR1, GABRR2, GABRR3, LAMB5, SLC6A11.

The expression data from all genes were pairwise correlated using Spearman correlations and the correlation matrix was clustered using the default unsupervised hierarchical clustering method implemented in *scipy* (29), based on Euclidean distances and estimating cluster proximity using the nearest point algorithm. Normalized expression values of all genes within a cluster were Z-standardized, averaged and correlated with the INS ALE-Z-map using *JuSpyce* (30) as described in the main methods section (partial Spearman correlations, adjusted for local gray matter volume). Resulting *p* values were FDR-corrected.

### 1.8 Dominance analysis on nuclear imaging and neuronal cell type data

Dominance analysis is designed to determine the relative contribution of each predictor in a multivariate regression model to the overall explained variance as an intuitive measure of predictor importance (31). It does so by calculating all subset models of a multivariate linear model, i.e., recalculating the regression analysis with all possible combinations of predictors. We implemented the method in *JuSpyce* to estimate three different dominance statistics of which we focused on two. *Total dominance* is the average contribution of a predictor to the total  $R^2$  across all subset models and can be interpreted as the amount of explained variance of a predictor relative to the total explained variance. For comparison, we further evaluated *individual dominance*, which is the  $R^2$  resulting from the univariate regression of a single predictor on the target variable and thus quantifies the amount of information explained by a predictor alone. As the main goal of this analysis was to quantify the amount of INS variance explained by PET and cell type maps, we submitted only those maps to dominance analysis that showed significant positive relationships to INS (FDR-corrected) in the previous analyses. PET data was atlas-wise parcellated, Z-standardized, and averaged across atlases using the same tracer. Neuronal cell type maps were generated by Z-standardizing each parcel-wise gene expression vector and calculating the average gene expression per cell type category. Parcel-wise gray matter volume was regressed out of the INS map and each predictor map before performing the regression analysis to align with the main correlation analyses.

### 1.9 Clustering and visualization of INS-associated GeneOntology categories

Results from GeneOntology (GO) gene-category enrichment analyses as obtained from *ABAnnotate* (32) were further clustered based on semantic similarity using *GO-Figure!* which was described in detail elsewhere (33, 34). Briefly, the dimensionality of the list of significantly associated GO terms is reduced by calculating pairwise semantic similarity scores based on (i) the distance of two GO terms within the directed acyclic organization of GO categories and (ii) the frequency of each GO term in a large database of genes (equaling “specificity” of each GO term).

Based on an arbitrary similarity threshold, GO terms are then grouped into clusters, a representative term is selected, and a multidimensional scaling algorithm is applied to generate a two-dimensional visualization. Here, we chose a liberal threshold of  $\geq .2$  to capture the overall biological functions of the identified GO clusters at the cost of specificity.

## 2 Supplementary Results

### 2.1 FMRI meta-analysis sensitivity analyses

#### 2.1.1 Influence of individual experiments

To estimate the influence of individual experiments on observed interpersonal neural synchrony (INS) clusters, we applied a jackknife approach and recalculated ALE analyses while iteratively excluding one experiment at a time.

Using these data, we estimated that 12 of 22 experiments contributed to the right temporoparietal junction (rTPJ) cluster, with a maximum contribution of 16% using the conservative threshold ( $p < .001$ ). In contrast, the right superior temporal and right insula clusters were driven by a lower number of studies with corresponding stronger contributions (Table S1). Accordingly, the spatial conjunction of all jackknife-derived thresholded maps proofed only the right temporoparietal junction (rTPJ) cluster as stable against the influence of individual experiments (Figure 2A).

#### 2.1.2 Risk of publication bias

Coordinate-based meta-analytic techniques, such as ALE, only allow for limited possibilities for structured bias assessment. However, to assess the robustness of ALE results against publication bias, calculation of the cluster-wise *fail-safe-N* was proposed (8). Adopting this method, we tested how many noise-experiments generated with characteristics similar to the original INS data we could add to the INS ALE analysis before the originally observed clusters were no longer significant.

Based on the inclusion of a minimum of 12 (the number of contributing experiments) and a maximum of 98 ( $\frac{12}{98+22} = 10\%$ ) of noise-experiments, we observed a fail-safe-N of 66 for the rTPJ cluster indicating that even if 66 “negative” experiments were not available for this analysis due to publication bias, we would still have observed spatial convergence of INS in the rTPJ. By contrast, the cluster in the right superior temporal gyrus did not survive the inclusion of the defined minimum of 12 additional noise-experiments.

### 2.2 fNIRS meta-analysis sensitivity analyses

#### 2.2.1 Alternative “fNIRS-Indices”

In addition to the “INS-to-all-channel-ratio” weighted by subject number, we also evaluated the ratio weighted by the number of experiments as well as the raw count of “INS-channels” as parcel-wise indices. We, however, emphasize that only the first index which is reported in the main paper incorporates all available information (presence of “INS-channels” in relation to the times a region was sampled and number of subjects)

Based on exact  $p$  values estimated from permutation of parcel-channel assignments, we observed significant results for left inferior-anterior prefrontal regions across all evaluated measures. Additionally, while evaluation of the channel ratio weighted by subjects indicated right temporal brain regions, results based on the INS channel count pointed to left temporal and parietal regions and based on the channel-ratio weighted by the number of experiments we found a significant right frontomedial parcel (Figure S5). Again, no  $p$  value survived FDR correction.

### **2.2.2 Modelling of spatial uncertainty**

To account for spatial uncertainty of fNIRS data in all fNIRS meta-analyses, we iteratively repeated the parcel-wise fNIRS meta-analysis as well as the ALE analysis with spatially randomized fNIRS coordinates (1 cm radius, 1,000 iterations) and assessed the percentages of iterations in which significant results at the original brain locations were observed.

In the fNIRS-only meta-analysis, we generally observed a high sensitivity of cluster significance towards randomization of parcel-coordinates. Especially the relatively small rTPJ parcel was highly sensitive, showing sub-threshold  $p$  values in only 7.9 % of iterations (*median*  $p = .28$ ) in the main evaluated fNIRS index. The highest consistency was found for the right inferior temporal parcel showing persistence in 48 % of iterations (*median*  $p = .052$ ). The overall results for all fNIRS indices are displayed in Table S6 and Figure S5. Concerning the combined fMRI-fNIRS ALE, coordinate randomization least affected the rTPJ and the left and right superior frontal clusters which emerged in 100 % of iterations, while the right middle frontal cluster was found in 89.1 % of iterations.

### **2.2.3 Restricted set of fNIRS experiments**

To confirm our fNIRS-results, we restricted the included experiments to a more conservative selection including only experiments explicitly contrasting interaction with control, rest, or randomization conditions and recalculated all analyses.

We observed a generally comparable pattern but in the parcellation-based fNIRS-only meta-analysis only prefrontal parcels showed significance for the “INS channel ratios” weighted by number of subjects or experiments while the fNIRS evaluation based on the raw INS channel count remained stable ( $p$  uncorrected; Figure S5). In the combined fMRI-fNIRS ALE, we mainly observed a reduction of cluster sizes (Tables S1 and S2).

## **2.3 MACM sensitivity analyses**

### **2.3.1 Controlling for baseline activation probability**

Specific coactivation likelihood estimation (SCALE) is an alternative meta-analytic connectivity modeling (MACM) algorithm controlling for the baseline probability of observing coactivation independent of the chosen seed (35). We calculated a SCALE analysis using all studies in the BrainMap database as baseline ( $N = 3,098$ ), to estimate the brain regions most uniquely coactivating with the rTPJ.

Here, we observed the left TPJ region, and to a lesser extent right insula, as specific functional connections of the rTPJ, further indicating the TPJs as hub regions of the observed INS-related network (Figure S3B).

### 2.3.2 Spatial alignment between INS-ALE and MACM activation patterns

To estimate whether the MACM network mirrors a brain-wide activation pattern that was already present in the original meta-analytic INS map beyond the rTPJ activation, we correlated the parcellated whole-brain maps derived from both analyses.

We observed a spatial alignment pattern driven by bilateral TPJs, insulae as well as dorsolateral prefrontal cortices, possibly indicating a role of these regions in INS but a lack of power to detect these as areas of spatial convergence in the main meta-analysis (Figure S3C).

## 2.4 Spatial relationships to established resting-state networks

We characterized the rTPJ cluster and the MACM network in respect to their spatial overlap with established brain-wide resting-state networks (15). We adopted a frequently used method characterizing the *absolute* and *relative distributions* of our target volumes across these networks (see method). We then permuted the input coordinates within all gray matter voxels (1,000 iterations), tested the resulting maps for meta-analytic convergence, and calculated absolute and relative distributions for each null map. From these null distributions, empirical  $p$  values were estimated.

The result is shown in Figure 2E. Looking at the absolute distributions, the rTPJ cluster was significantly associated with the default mode network ( $p = .001, q = .002$ ), the dorsal attention ( $p = .001, q = .002$ ), and the ventral attention network ( $p = .026, q = .040$ ). The MACM network showed a widespread pattern (default mode, dorsal attention, ventral attention, frontoparietal, and somatomotor network:  $p = .001, q = .002$ ). Concerning the relative distributions, the rTPJ cluster did not show significant associations while, in confirmation of the relationship between INS and attention networks, associations significant at an uncorrected alpha-level were observed between the MACM network and the ventral ( $p = .004, q = .056$ ) and dorsal ( $p = .047, q = .243$ ) attention networks.

## 2.5 Neurotransmitter-associations sensitivity analyses

### 2.5.1 GABA<sub>A</sub>-related mRNA expression

To validate the strong association found to the GABA<sub>A</sub> receptor, and further clarify which subtype of GABA<sub>A</sub> receptors may drive the association, we collected a list of GABA-related genes (27, 28), clustered these according to their spatial co-expression profiles (Figure S7A), and assessed spatial correlation patterns between INS and cluster-wise mRNA expression data.

Two GABA gene clusters were significantly associated with INS ( $Z = .43, p = .007, q = .014$  and  $Z = .45, p = .005, q = .014$ ). In line with prior data on the molecular target of the applied GABA<sub>A</sub> tracer (27), one of these clusters comprised the  $\alpha 1$ -GABA<sub>A</sub> receptor subunit (GABRA1) and parvalbumin (PVALB), of which the latter is considered a marker of fast-spiking parvalbumin-expressing interneurons, the largest group of cortical inhibitory neurons (36) (Figure S7B, clusters 2 and 3; Table S8).

### 3 Supplementary References

1. J. L. Lancaster, D. Tordesillas-Gutiérrez, M. Martinez, F. Salinas, A. Evans, K. Zilles, J. C. Mazziotta, P. T. Fox, Bias between MNI and Talairach coordinates analyzed using the ICBM-152 brain template. *Hum. Brain Mapp.* **28**, 1194–1205 (2007).
2. D. Tsuzuki, V. Jurcak, A. K. Singh, M. Okamoto, E. Watanabe, I. Dan, Virtual spatial registration of stand-alone fNIRS data to MNI space. *NeuroImage*. **34**, 1506–1518 (2007).
3. X. Cui, D. M. Bryant, A. L. Reiss, NIRS-based hyperscanning reveals increased interpersonal coherence in superior frontal cortex during cooperation. *NeuroImage*. **59**, 2430–2437 (2012).
4. J. Liu, R. Zhang, B. Geng, T. Zhang, D. Yuan, S. Otani, X. Li, Interplay between prior knowledge and communication mode on teaching effectiveness: Interpersonal neural synchronization as a neural marker. *NeuroImage*. **193**, 93–102 (2019).
5. K. Lu, T. Yu, N. Hao, Creating while taking turns, the choice to unlocking group creative potential. *NeuroImage*. **219**, 117025 (2020).
6. R. M. Ball, thesis (2011).
7. C. R. Harrison, K. M. Robinette, “CAESAR: Summary statistics for the adult population (ages 18-65) of the United States of America” (AIR FORCE RESEARCH LAB WRIGHT-PATTERSON AFB OH HUMAN EFFECTIVENESS DIRECTORATE, 2002).
8. F. Acar, R. Seurinck, S. Eickhoff, B. Moerkerke, Assessing robustness against potential publication bias in Activation Likelihood Estimation (ALE) meta-analyses for fMRI. *PLOS ONE*. **13**, e0208177 (2018).
9. R. J. Cooper, M. Caffini, J. Dubb, Q. Fang, A. Custo, D. Tsuzuki, B. Fischl, W. Wells, I. Dan, D. A. Boas, Validating atlas-guided DOT: A comparison of diffuse optical tomography informed by atlas and subject-specific anatomies. *NeuroImage*. **62**, 1999–2006 (2012).
10. D. Tsuzuki, I. Dan, Spatial registration for functional near-infrared spectroscopy: From channel position on the scalp to cortical location in individual and group analyses. *NeuroImage*. **85**, 92–103 (2014).
11. D. C. Van Essen, S. M. Smith, D. M. Barch, T. E. J. Behrens, E. Yacoub, K. Ugurbil, The WU-Minn Human Connectome Project: An overview. *NeuroImage*. **80**, 62–79 (2013).
12. WU-Minn Consortium Human Connectome Project, WU-Minn HCP 900 Subjects Data Release Reference Manual (2015), (available at <https://www.humanconnectome.org/study/hcp-young-adult/document/900-subjects-data-release>).
13. S. Whitfield-Gabrieli, A. Nieto-Castanon, Conn: a functional connectivity toolbox for correlated and anticorrelated brain networks. *Brain connectivity*. **2**, 125–141 (2012).
14. K. Murphy, R. M. Birn, D. A. Handwerker, T. B. Jones, P. A. Bandettini, The impact of global signal regression on resting state correlations: Are anti-correlated networks introduced? *NeuroImage*. **44**, 893–905 (2009).
15. B. T. T. Yeo, F. M. Krienen, J. Sepulcre, M. R. Sabuncu, D. Lashkari, M. Hollinshead, J. L. Roffman, J. W. Smoller, L. Zöllei, J. R. Polimeni, B. Fischl, H. Liu, R. L. Buckner, The organization of the human cerebral cortex estimated by intrinsic functional connectivity. *Journal of Neurophysiology*. **106**, 1125–1165 (2011).
16. E. Y. Choi, B. T. T. Yeo, R. L. Buckner, The organization of the human striatum estimated by intrinsic functional connectivity. *J Neurophysiol*. **108**, 2242–2263 (2012).
17. B. T. T. Yeo, Thalamic resting-state networks. *Twitter* (2020), (available at <https://twitter.com/bttyeo/status/1248992127985434624>).
18. T. Chen, B. Becker, J. Camilleri, L. Wang, S. Yu, S. B. Eickhoff, C. Feng, A domain-general brain network underlying emotional and cognitive interference processing: evidence from coordinate-based and functional connectivity meta-analyses. *Brain Struct Funct*. **223**, 3813–3840 (2018).
19. M. J. Hawrylycz, E. S. Lein, A. L. Guillozet-Bongaarts, E. H. Shen, L. Ng, J. A. Miller, L. N. van de Lagemaat, K. A. Smith, A. Ebbert, Z. L. Riley, C. Abajian, C. F. Beckmann, A. Bernard, D. Bertagnolli, A. F. Boe, P. M. Cartagena, M. M. Chakravarty, M. Chapin, J. Chong, R. A. Dalley, B. D. Daly, C. Dang, S. Datta, N. Dec, T. A. Dolbeare, V. Faber, D. R. Fowler, J. Goldy, B. W. Gregor, Z. Haradon, D. R. Haynor, J. G. Hohmann, S. Horvath, R. E. Howard, A. Jeromin, J. M. Jochim, M. Kinnunen, C. Lau, E. T. Lazars, C. Lee, T. A. Lemon, L. Li, Y. Li, J. A. Morris, C. C. Overly, P. D. Parker, S. E. Parry, M. Reding, J. J. Royall, J. Schulkin, P. A. Sequeira, C. R. Slaughterbeck, S. C. Smith, A. J. Sodt, S. M. Sunkin, B. E. Swanson, M. P. Pawter, D. Williams, P. Wohnoutka, H. R. Zielke, D. H. Geschwind, P. R. Hof, S. M. Smith, C. Koch, S. G. N. Grant, A. R. Jones, An anatomically comprehensive atlas of the adult human brain transcriptome. *Nature*. **489**, 391–399 (2012).
20. R. D. Markello, A. Arnatkeviciute, J.-B. Poline, B. D. Fulcher, A. Fornito, B. Misic, Standardizing workflows in imaging transcriptomics with the abagen toolbox. *eLife*. **10**, e72129 (2021).
21. A. Schaefer, R. Kong, E. M. Gordon, T. O. Laumann, X.-N. Zuo, A. J. Holmes, S. B. Eickhoff, B. T. T. Yeo, Local-Global Parcellation of the Human Cerebral Cortex from Intrinsic Functional Connectivity MRI. *Cereb Cortex*. **28**, 3095–3114 (2018).
22. Y. Tian, D. S. Margulies, M. Breakspear, A. Zalesky, Topographic organization of the human subcortex unveiled with functional connectivity gradients. *Nat Neurosci*. **23**, 1421–1432 (2020).
23. A. Arnatkeviciute, B. D. Fulcher, A. Fornito, A practical guide to linking brain-wide gene expression and neuroimaging data. *NeuroImage*. **189**, 353–367 (2019).
24. J. Quackenbush, Microarray data normalization and transformation. *Nat Genet*. **32 Suppl**, 496–501 (2002).
25. M. Hawrylycz, J. A. Miller, V. Menon, D. Feng, T. Dolbeare, A. L. Guillozet-Bongaarts, A. G. Jegga, B. J. Aronow, C.-K. Lee, A. Bernard, M. F. Glasser, D. L. Dierker, J. Menche, A. Szafer, F. Collman, P. Grange, K. A. Berman, S. Mihalas, Z. Yao, L. Stewart, A.-L. Barabási, J. Schulkin, J. Phillips, L. Ng, C. Dang, D. R. Haynor, A. Jones, D. C. Van Essen, C. Koch, E. Lein, Canonical genetic signatures of the adult human brain. *Nat Neurosci*. **18**, 1832–1844 (2015).
26. B. D. Fulcher, M. A. Little, N. S. Jones, Highly comparative time-series analysis: the empirical structure of time series and their methods. *Journal of The Royal Society Interface*. **10**, 20130048 (2013).
27. P. B. Lukow, D. Martins, M. Veronese, A. C. Vernon, P. McGuire, F. E. Turkheimer, G. Modinos, Cellular and molecular signatures of in vivo GABAergic neurotransmission in the human brain (2021), , doi:10.1101/2021.06.17.448812.
28. L. Martini, R. Bardini, S. D. Carlo, Meta-Analysis of cortical inhibitory interneurons markers landscape and their performances in scRNA-seq studies (2021), , doi:10.1101/2021.11.03.467049.
29. P. Virtanen, R. Gommers, T. E. Oliphant, M. Haberland, T. Reddy, D. Cournapeau, E. Burovski, P. Peterson, W. Weckesser, J. Bright, S. J. van der Walt, M. Brett, J. Wilson, K. J. Millman, N. Mayorov, A. R. J. Nelson, E. Jones, R. Kern, E. Larson, C. J. Carey, Í. Polat, Y. Feng, E. W. Moore, J. VanderPlas, D. Laxalde, J. Perktold, R. Cimrman, I. Henriksen, E. A. Quintero, C. R. Harris, A. M. Archibald, A. H. Ribeiro, F. Pedregosa, P. van Mulbregt, SciPy 1.0: fundamental algorithms for scientific computing in Python. *Nat Methods*. **17**, 261–272 (2020).
30. L. D. Lotter, J. Dukart, JuSpyce - a toolbox for flexible assessment of spatial associations between brain images (2022), , doi:10.5281/zenodo.6884932.

31. R. Azen, D. V. Budescu, The dominance analysis approach for comparing predictors in multiple regression. *Psychol Methods*. **8**, 129–148 (2003).
32. L. D. Lotter, J. Dukart, B. D. Fulcher, ABAnnotate: A toolbox for ensemble-based multimodal gene-category enrichment analysis of human neuroimaging data (2022), doi:10.5281/zenodo.6463329.
33. M. J. M. F. Reijnders, R. M. Waterhouse, Summary Visualizations of Gene Ontology Terms With GO-Figure! *Frontiers in Bioinformatics*. **1** (2021) (available at <https://www.frontiersin.org/article/10.3389/fbinf.2021.638255>).
34. C. Pesquita, "Semantic Similarity in the Gene Ontology" in *The Gene Ontology Handbook*, C. Dessimoz, N. Škunca, Eds. (Springer, New York, NY, 2017; [https://doi.org/10.1007/978-1-4939-3743-1\\_12](https://doi.org/10.1007/978-1-4939-3743-1_12)), *Methods in Molecular Biology*, pp. 161–173.
35. R. Langner, C. Rottschy, A. R. Laird, P. T. Fox, S. B. Eickhoff, Meta-analytic connectivity modeling revisited: Controlling for activation base rates. *NeuroImage*. **99**, 559–570 (2014).
36. R. Tremblay, S. Lee, B. Rudy, GABAergic Interneurons in the Neocortex: From Cellular Properties to Circuits. *Neuron*. **91**, 260–292 (2016).
37. E. Bilek, M. Ruf, A. Schäfer, C. Akdeniz, V. D. Calhoun, C. Schmah, C. Demanuele, H. Tost, P. Kirsch, A. Meyer-Lindenberg, Information flow between interacting human brains: Identification, validation, and relationship to social expertise. *Proc Natl Acad Sci USA*. **112**, 5207–5212 (2015).
38. I. Gordon, S. Wallot, Y. Berson, Group-level physiological synchrony and individual-level anxiety predict positive affective behaviors during a group decision-making task. *Psychophysiology*. **58**, e13857 (2021).
39. T. Koike, H. C. Tanabe, S. Okazaki, E. Nakagawa, A. T. Sasaki, K. Shimada, S. K. Sugawara, H. K. Takahashi, K. Yoshihara, J. Bosch-Bayard, N. Sadato, Neural substrates of shared attention as social memory: A hyperscanning functional magnetic resonance imaging study. *NeuroImage*. **125**, 401–412 (2016).
40. T. Koike, M. Sumiya, E. Nakagawa, S. Okazaki, N. Sadato, What Makes Eye Contact Special? Neural Substrates of On-Line Mutual Eye-Gaze: A Hyperscanning fMRI Study. *eNeuro*. **6** (2019), doi:10.1523/ENEURO.0284-18.2019.
41. T. Koike, H. C. Tanabe, S. Adachi-Abe, S. Okazaki, E. Nakagawa, A. T. Sasaki, K. Shimada, S. K. Sugawara, H. K. Takahashi, K. Yoshihara, N. Sadato, Role of the right anterior insular cortex in joint attention-related identification with a partner. *Soc Cogn Affect Neurosci*. **14**, 1131–1145 (2019).
42. K. Miyata, T. Koike, E. Nakagawa, T. Harada, M. Sumiya, T. Yamamoto, N. Sadato, Neural substrates for sharing intention in action during face-to-face imitation. *NeuroImage*. **233**, 117916 (2021).
43. D. N. Saito, H. C. Tanabe, K. Izuma, M. J. Hayashi, Y. Morito, H. Komeda, H. Uchiyama, H. Kosaka, H. Okazawa, Y. Fujibayashi, N. Sadato, "Stay Tuned": Inter-Individual Neural Synchronization During Mutual Gaze and Joint Attention. *Front. Integr. Neurosci*. **4** (2010), doi:10.3389/fnint.2010.00127.
44. M. Salazar, D. J. Shaw, M. Gajdoš, R. Mareček, K. Czekóová, M. Mikl, M. Brázdil, You took the words right out of my mouth: Dual-fMRI reveals intra- and inter-personal neural processes supporting verbal interaction. *NeuroImage*. **228**, 117697 (2021).
45. D. J. Shaw, K. Czekóová, R. Staněk, R. Mareček, T. Urbánek, J. Špalek, L. Kopečková, J. Řezáč, M. Brázdil, A dual-fMRI investigation of the iterated Ultimatum Game reveals that reciprocal behaviour is associated with neural alignment. *Scientific Reports*. **8**, 10896 (2018).
46. D. J. Shaw, K. Czekóová, B. Špiláková, M. Salazar, P. Řezáč, V. Kurečková, P. Zámečník, M. Brázdil, A neuroscientific evaluation of driver rehabilitation: Functional neuroimaging demonstrates the effectiveness of empathy induction in altering brain responses during social information processing. *PLoS One*. **15**, e0232222 (2020).
47. K. Spiegelhalter, S. Ohlendorf, W. Regen, B. Feige, L. Tebartz van Elst, C. Weiller, J. Hennig, M. Berger, O. Tüscher, Interindividual synchronization of brain activity during live verbal communication. *Behavioural Brain Research*. **258**, 75–79 (2014).
48. B. Špiláková, D. J. Shaw, K. Czekóová, R. Mareček, M. Brázdil, Getting into sync: Data-driven analyses reveal patterns of neural coupling that distinguish among different social exchanges. *Human Brain Mapping*. **41**, 1072–1083 (2020).
49. L.-S. Wang, J.-T. Cheng, I.-J. Hsu, S. Liou, C.-C. Kung, D.-Y. Chen, M.-H. Weng, Distinct cerebral coherence in task-based fMRI hyperscanning: cooperation versus competition (2022), doi:10.1093/cercor/bhac075.
50. H. Xie, I. I. Karipidis, A. Howell, M. Schreier, K. E. Sheau, M. K. Manchanda, R. Ayub, G. H. Glover, M. Jung, A. L. Reiss, M. Saggat, Finding the neural correlates of collaboration using a three-person fMRI hyperscanning paradigm. *PNAS*. **117**, 23066–23072 (2020).
51. A. Yoshioka, H. C. Tanabe, M. Sumiya, E. Nakagawa, S. Okazaki, T. Koike, N. Sadato, Neural substrates of shared visual experiences: a hyperscanning fMRI study. *Social Cognitive and Affective Neuroscience*. **16**, 1264–1275 (2021).
52. S. Anders, J. Heinzle, N. Weiskopf, T. Ethofer, J.-D. Haynes, Flow of affective information between communicating brains. *NeuroImage*. **54**, 439–446 (2011).
53. S. Dikker, L. J. Silbert, U. Hasson, J. D. Zevin, On the Same Wavelength: Predictable Language Enhances Speaker–Listener Brain-to-Brain Synchrony in Posterior Superior Temporal Gyrus. *J. Neurosci*. **34**, 6267–6272 (2014).
54. K. Kistorz, V. L. Flanagan, S. Glasauer, Synchronization between instructor and observer when learning a complex bimanual skill. *NeuroImage*. **216**, 116659 (2020).
55. L. Liu, X. Ding, H. Li, Q. Zhou, D. Gao, C. Lu, G. Ding, Reduced listener–speaker neural coupling underlies speech understanding difficulty in older adults. *Brain Struct Funct*. **226**, 1571–1584 (2021).
56. L. Liu, H. Li, Z. Ren, Q. Zhou, Y. Zhang, C. Lu, J. Qiu, H. Chen, G. Ding, The "Two-Brain" Approach Reveals the Active Role of Task-Deactivated Default Mode Network in Speech Comprehension. *Cerebral Cortex* (2022), doi:10.1093/cercor/bhab521.
57. L. J. Silbert, C. J. Honey, E. Simony, D. Poeppel, U. Hasson, Coupled neural systems underlie the production and comprehension of naturalistic narrative speech. *PNAS*. **111**, E4687–E4696 (2014).
58. D. Smirnov, H. Saarimäki, E. Glerean, R. Hari, M. Sams, L. Nummenmaa, Emotions amplify speaker–listener neural alignment. *Human Brain Mapping*. **40**, 4777–4788 (2019).
59. G. J. Stephens, L. J. Silbert, U. Hasson, Speaker–listener neural coupling underlies successful communication. *Proceedings of the National Academy of Sciences*. **107**, 14425–14430 (2010).
60. M. Balconi, G. Fronza, M. E. Vanutelli, Donate or receive? Social hyperscanning application with fNIRS. *Curr Psychol*. **38**, 991–1002 (2019).
61. M. Balconi, L. Pezard, J.-L. Nandrino, M. E. Vanutelli, Two is better than one: The effects of strategic cooperation on intra- and inter-brain connectivity by fNIRS. *PLOS ONE*. **12**, e0187652 (2017).
62. M. Balconi, M. E. Vanutelli, L. Gatti, Functional brain connectivity when cooperation fails. *Brain Cogn*. **123**, 65–73 (2018).
63. R. Cañigueral, X. Zhang, J. A. Noah, I. Tachtsidis, A. F. de C. Hamilton, J. Hirsch, Facial and neural mechanisms during interactive disclosure of biographical information. *NeuroImage*. **226**, 117572 (2021).
64. M. Chen, T. Zhang, R. Zhang, N. Wang, Q. Yin, Y. Li, J. Liu, T. Liu, X. Li, Neural alignment during face-to-face spontaneous deception: Does gender make a difference? *Human Brain Mapping*. **41**, 4964–4981 (2020).
65. X. Cheng, X. Li, Y. Hu, Synchronous brain activity during cooperative exchange depends on gender of partner: A fNIRS-based hyperscanning study. *Hum Brain Mapp*. **36**, 2039–2048 (2015).

66. X. Cheng, Y. Pan, Y. Hu, Y. Hu, Coordination Elicits Synchronous Brain Activity Between Co-actors: Frequency Ratio Matters. *Frontiers in Neuroscience*. **13** (2019) (available at <https://www.frontiersin.org/article/10.3389/fnins.2019.01071>).
67. X. Cheng, Y. Zhu, Y. Hu, X. Zhou, Y. Pan, Y. Hu, Integration of social status and trust through interpersonal brain synchronization. *NeuroImage*. **246**, 118777 (2022).
68. H. Duan, T. Yang, X. Wang, Y. Kan, H. Zhao, Y. Li, W. Hu, Is the creativity of lovers better? A behavioral and functional near-infrared spectroscopy hyperscanning study. *Current Psychology*. **41**, 41–54 (2022).
69. X. Feng, B. Sun, C. Chen, W. Li, Y. Wang, W. Zhang, W. Xiao, Y. Shao, Self–other overlap and interpersonal neural synchronization serially mediate the effect of behavioral synchronization on prosociality. *Social Cognitive and Affective Neuroscience*. **15**, 203–214 (2020).
70. G. Fronza, M. Balconi, The effect of interbrain synchronization in gesture observation: A fNIRS study. *Brain and Behavior*. **10**, e01663 (2020).
71. Y. Hu, Y. Hu, X. Li, Y. Pan, X. Cheng, Brain-to-brain synchronization across two persons predicts mutual prosociality. *Social Cognitive and Affective Neuroscience*. **12**, 1835–1844 (2017).
72. T. Koide, S. Shimada, Cheering Enhances Inter-Brain Synchronization Between Sensorimotor Areas of Player and Observer. *Japanese Psychological Research*. **60**, 265–275 (2018).
73. L. Li, H. Wang, H. Luo, X. Zhang, R. Zhang, X. Li, Interpersonal Neural Synchronization During Cooperative Behavior of Basketball Players: A fNIRS-Based Hyperscanning Study. *Frontiers in Human Neuroscience*. **14** (2020) (available at <https://www.frontiersin.org/article/10.3389/fnhum.2020.00169>).
74. Y. Li, M. Chen, R. Zhang, X. Li, Experiencing happiness together facilitates dyadic coordination through the enhanced interpersonal neural synchronization. *Social Cognitive and Affective Neuroscience*, nsab114 (2021).
75. Y. Li, R. Chen, O. Turel, T. Feng, C.-Z. Zhu, Q. He, Dyad sex composition effect on inter-brain synchronization in face-to-face cooperation. *Brain Imaging and Behavior*. **15**, 1667–1675 (2021).
76. J. Liu, R. Zhang, B. Geng, T. Zhang, D. Yuan, S. Otani, X. Li, Interplay between prior knowledge and communication mode on teaching effectiveness: Interpersonal neural synchronization as a neural marker. *NeuroImage*. **193**, 93–102 (2019).
77. N. Liu, C. Mok, E. E. Witt, A. H. Pradhan, J. E. Chen, A. L. Reiss, NIRS-Based Hyperscanning Reveals Inter-brain Neural Synchronization during Cooperative Jenga Game with Face-to-Face Communication. *Front. Hum. Neurosci.* **10** (2016), doi:10.3389/fnhum.2016.00082.
78. T. Liu, G. Saito, C. Lin, H. Saito, Inter-brain network underlying turn-based cooperation and competition: A hyperscanning study using near-infrared spectroscopy. *Scientific Reports*. **7**, 8684 (2017).
79. W. Liu, H. P. Branigan, L. Zheng, Y. Long, X. Bai, K. Li, H. Zhao, S. Zhou, M. J. Pickering, C. Lu, Shared neural representations of syntax during online dyadic communication. *NeuroImage*. **198**, 63–72 (2019).
80. Y. Long, C. Chen, K. Wu, S. Zhou, F. Zhou, L. Zheng, H. Zhao, Y. Zhai, C. Lu, Interpersonal Conflict Increases Interpersonal Neural Synchronization in Romantic Couples. *Cerebral Cortex*, bhab413 (2021).
81. Y. Long, L. Zheng, H. Zhao, S. Zhou, Y. Zhai, C. Lu, Interpersonal Neural Synchronization during Interpersonal Touch Underlies Affiliative Pair Bonding between Romantic Couples. *Cerebral Cortex*. **31**, 1647–1659 (2021).
82. K. Lu, N. Hao, When do we fall in neural synchrony with others? *Social Cognitive and Affective Neuroscience*. **14**, 253–261 (2019).
83. K. Lu, X. Qiao, N. Hao, Praising or keeping silent on partner’s ideas: Leading brainstorming in particular ways. *Neuropsychologia*. **124**, 19–30 (2019).
84. K. Lu, J. Teng, N. Hao, Gender of partner affects the interaction pattern during group creative idea generation. *Exp Brain Res*. **238**, 1157–1168 (2020).
85. K. Lu, H. Xue, T. Nozawa, N. Hao, Cooperation Makes a Group be More Creative. *Cerebral Cortex*. **29**, 3457–3470 (2019).
86. T. Nozawa, K. Sakaki, S. Ikeda, H. Jeong, S. Yamazaki, K. H. dos S. Kawata, N. Y. dos S. Kawata, Y. Sasaki, K. Kulason, K. Hirano, Y. Miyake, R. Kawashima, Prior physical synchrony enhances rapport and inter-brain synchronization during subsequent educational communication. *Sci Rep*. **9**, 12747 (2019).
87. T. Nozawa, Y. Sasaki, K. Sakaki, R. Yokoyama, R. Kawashima, Interpersonal frontopolar neural synchronization in group communication: An exploration toward fNIRS hyperscanning of natural interactions. *NeuroImage*. **133**, 484–497 (2016).
88. Y. Pan, X. Cheng, Z. Zhang, X. Li, Y. Hu, Cooperation in lovers: An fNIRS-based hyperscanning study. *Hum Brain Mapp*. **38**, 831–841 (2017).
89. Y. Pan, S. Dikker, P. Goldstein, Y. Zhu, C. Yang, Y. Hu, Instructor-learner brain coupling discriminates between instructional approaches and predicts learning. *NeuroImage*. **211**, 116657 (2020).
90. Y. Pan, C. Guyon, G. Borragán, Y. Hu, P. Peigneux, Interpersonal brain synchronization with instructor compensates for learner’s sleep deprivation in interactive learning. *Biochemical Pharmacology*. **191**, 114111 (2021).
91. Y. Pan, G. Novembre, B. Song, X. Li, Y. Hu, Interpersonal synchronization of inferior frontal cortices tracks social interactive learning of a song. *NeuroImage*. **183**, 280–290 (2018).
92. P. Pinti, A. Devoto, I. Greenhalgh, I. Tachtsidis, P. W. Burgess, A. F. de C Hamilton, The role of anterior prefrontal cortex (area 10) in face-to-face deception measured with fNIRS. *Social Cognitive and Affective Neuroscience*. **16**, 129–142 (2021).
93. L. E. Quiñones-Camacho, F. A. Fishburn, K. Belardi, D. L. Williams, T. J. Huppert, S. B. Perlman, Dysfunction in interpersonal neural synchronization as a mechanism for social impairment in autism spectrum disorder. *Autism Research*. **14**, 1585–1596 (2021).
94. B. Sun, W. Xiao, X. Feng, Y. Shao, W. Zhang, W. Li, Behavioral and brain synchronization differences between expert and novice teachers when collaborating with students. *Brain and Cognition*. **139**, 105513 (2020).
95. B. Sun, W. Xiao, S. Lin, Y. Shao, W. Li, W. Zhang, Cooperation with partners of differing social experience: An fNIRS-based hyperscanning study. *Brain and Cognition*. **154**, 105803 (2021).
96. H. Tang, X. Mai, S. Wang, C. Zhu, F. Krueger, C. Liu, Interpersonal brain synchronization in the right temporo-parietal junction during face-to-face economic exchange. *Social Cognitive and Affective Neuroscience*. **11**, 23–32 (2016).
97. C. Wang, T. Zhang, Z. Shan, J. Liu, D. Yuan, X. Li, Dynamic interpersonal neural synchronization underlying pain-induced cooperation in females. *Hum Brain Mapp*. **40**, 3222–3232 (2019).
98. H. Xue, K. Lu, N. Hao, Cooperation makes two less-creative individuals turn into a highly-creative pair. *NeuroImage*. **172**, 527–537 (2018).
99. J. Yang, H. Zhang, J. Ni, C. K. W. De Dreu, Y. Ma, Within-group synchronization in the prefrontal cortex associates with intergroup conflict. *Nat Neurosci*. **23**, 754–760 (2020).
100. M. Zhang, H. Jia, G. Wang, Interbrain Synchrony of Team Collaborative Decision-Making: An fNIRS Hyperscanning Study. *Frontiers in Human Neuroscience*. **15** (2021) (available at <https://www.frontiersin.org/article/10.3389/fnhum.2021.702959>).
101. M. Zhang, H. Jia, M. Zheng, Interbrain Synchrony in the Expectation of Cooperation Behavior: A Hyperscanning Study Using Functional Near-Infrared Spectroscopy. *Frontiers in Psychology*. **11** (2020) (available at <https://www.frontiersin.org/article/10.3389/fpsyg.2020.542093>).



102. M. Zhang, H. Jia, M. Zheng, T. Liu, Group decision-making behavior in social dilemmas: Inter-brain synchrony and the predictive role of personality traits. *Personality and Individual Differences*. **168**, 110315 (2021).
103. M. Zhang, T. Liu, M. Pelowski, D. Yu, Gender difference in spontaneous deception: A hyperscanning study using functional near-infrared spectroscopy. *Scientific Reports*. **7**, 7508 (2017).
104. M. Zhang, T. Liu, M. Pelowski, H. Jia, D. Yu, Social risky decision-making reveals gender differences in the TPJ: A hyperscanning study using functional near-infrared spectroscopy. *Brain Cogn*. **119**, 54–63 (2017).
105. R. Zhang, X. Zhou, D. Feng, D. Yuan, S. Li, C. Lu, X. Li, Effects of acute psychosocial stress on interpersonal cooperation and competition in young women. *Brain and Cognition*. **151**, 105738 (2021).
106. Y. Zhang, T. Meng, Y. Yang, Y. Hu, Experience-Dependent Counselor-Client Brain Synchronization during Psychological Counseling. *eNeuro*. **7** (2020), doi:10.1523/ENEURO.0236-20.2020.
107. H. Zhao, Y. Li, Y. Wang, X. Wang, Y. Kan, T. Yang, W. Hu, H. Duan, Acute stress makes women's group decisions more rational: A functional near-infrared spectroscopy (fNIRS)-based hyperscanning study. *Journal of Neuroscience, Psychology, and Economics*. **14**, 20–35 (2021).
108. Y. Zhao, R.-N. Dai, X. Xiao, Z. Zhang, L. Duan, Z. Li, C.-Z. Zhu, Independent component analysis-based source-level hyperlink analysis for two-person neuroscience studies. *JBO*. **22**, 027004 (2017).
109. L. Zheng, C. Chen, W. Liu, Y. Long, H. Zhao, X. Bai, Z. Zhang, Z. Han, L. Liu, T. Guo, B. Chen, G. Ding, C. Lu, Enhancement of teaching outcome through neural prediction of the students' knowledge state. *Hum Brain Mapp*. **39**, 3046–3057 (2018).
110. L. Zheng, W. Liu, Y. Long, Y. Zhai, H. Zhao, X. Bai, S. Zhou, K. Li, H. Zhang, L. Liu, T. Guo, G. Ding, C. Lu, Affiliative bonding between teachers and students through interpersonal synchronisation in brain activity. *Social Cognitive and Affective Neuroscience*. **15**, 97–109 (2020).
111. Y. Zhu, V. Leong, Y. Hou, D. Zhang, Y. Pan, Y. Hu, Instructor-learner neural synchronization during elaborated feedback predicts learning transfer. *Journal of Educational Psychology* (2021), doi:10.1037/edu0000707.
112. Y. Hou, B. Song, Y. Hu, Y. Pan, Y. Hu, The averaged inter-brain coherence between the audience and a violinist predicts the popularity of violin performance. *NeuroImage*. **211**, 116655 (2020).
113. Z. Li, J. Li, B. Hong, G. Nolte, A. K. Engel, D. Zhang, Speaker-Listener Neural Coupling Reveals an Adaptive Mechanism for Speech Comprehension in a Noisy Environment. *Cerebral Cortex*. **31**, 4719–4729 (2021).
114. Y. Liu, E. A. Piazza, E. Simony, P. A. Shewokis, B. Onaral, U. Hasson, H. Ayaz, Measuring speaker-listener neural coupling with functional near infrared spectroscopy. *Sci Rep*. **7**, 43293 (2017).
115. C. M. Aasted, M. A. Yücel, R. J. Cooper, J. Dubb, D. Tsuzuki, L. Becerra, M. P. Petkov, D. Borsook, I. Dan, D. A. Boas, Anatomical guidance for functional near-infrared spectroscopy: AtlasViewer tutorial. *NPh*. **2**, 020801 (2015).
116. M. P. Notter, D. Gale, P. Herholz, R. Markello, M.-L. Notter-Bielsler, K. Whitaker, AtlasReader: A Python package to generate coordinate tables, region labels, and informative figures from statistical MRI images. *Journal of Open Source Software*. **4**, 1257 (2019).
117. E. T. Rolls, C.-C. Huang, C.-P. Lin, J. Feng, M. Joliot, Automated anatomical labelling atlas 3. *NeuroImage*. **206**, 116189 (2020).
118. M. Savli, A. Bauer, M. Mitterhauser, Y.-S. Ding, A. Hahn, T. Kroll, A. Neumeister, D. Haeusler, J. Ungersboeck, S. Henry, S. A. Isfahani, F. Rattay, W. Wadsak, S. Kasper, R. Lanzenberger, Normative database of the serotonergic system in healthy subjects using multi-tracer PET. *Neuroimage*. **63**, 447–459 (2012).
119. J.-D. Gallezot, N. Nabulsi, A. Neumeister, B. Planeta-Wilson, W. A. Williams, T. Singhal, S. Kim, R. P. Maguire, T. McCarthy, J. J. Frost, Y. Huang, Y.-S. Ding, R. E. Carson, Kinetic modeling of the serotonin 5-HT(1B) receptor radioligand [(11C)P943] in humans. *J Cereb Blood Flow Metab*. **30**, 196–210 (2010).
120. V. Beliveau, M. Ganz, L. Feng, B. Ozenne, L. Højgaard, P. M. Fisher, C. Svarer, D. N. Greve, G. M. Knudsen, A High-Resolution In Vivo Atlas of the Human Brain's Serotonin System. *J Neurosci*. **37**, 120–128 (2017).
121. R. Radhakrishnan, N. Nabulsi, E. Gaiser, J.-D. Gallezot, S. Henry, B. Planeta, S. Lin, J. Ropchan, W. Williams, E. Morris, D. C. D'Souza, Y. Huang, R. E. Carson, D. Matuskey, Age-Related Change in 5-HT6 Receptor Availability in Healthy Male Volunteers Measured with 11C-GSK215083 PET. *J Nucl Med*. **59**, 1445–1450 (2018).
122. F. J. G. Gómez, I. Huertas, J. A. L. Ramírez, D. G. Solís, Elaboración de una plantilla de SPM para la normalización de imágenes de PET con 18F-DOPA. *Imagen Diagnóstica*. **9**, 23–25 (2018).
123. S. Kaller, M. Rullmann, M. Patt, G.-A. Becker, J. Luthardt, J. Girbardt, P. M. Meyer, P. Werner, H. Barthel, A. Bresch, T. H. Fritz, S. Hesse, O. Sabri, Test-retest measurements of dopamine D1-type receptors using simultaneous PET/MRI imaging. *Eur J Nucl Med Mol Imaging*. **44**, 1025–1032 (2017).
124. C. M. Sandiego, J.-D. Gallezot, K. Lim, J. Ropchan, S. Lin, H. Gao, E. D. Morris, K. P. Cosgrove, Reference region modeling approaches for amphetamine challenge studies with [11C]FLB 457 and PET. *J Cereb Blood Flow Metab*. **35**, 623–629 (2015).
125. C. T. Smith, J. L. Crawford, L. C. Dang, K. L. Seaman, M. D. San Juan, A. Vijay, D. T. Katz, D. Matuskey, R. L. Cowan, E. D. Morris, D. H. Zald, G. R. Samanez-Larkin, Partial-volume correction increases estimated dopamine D2-like receptor binding potential and reduces adult age differences. *J Cereb Blood Flow Metab*. **39**, 822–833 (2019).
126. J. Dukart, Š. Holiga, C. Chatham, P. Hawkins, A. Forsyth, R. McMillan, J. Myers, A. R. Lingford-Hughes, D. J. Nutt, E. Merlo-Pich, C. Risterucci, L. Boak, D. Umbricht, S. Schobel, T. Liu, M. A. Mehta, F. O. Zelaya, S. C. Williams, G. Brown, M. Paulus, G. D. Honey, S. Muthukumaraswamy, J. Hipp, A. Bertolino, F. Sambataro, Cerebral blood flow predicts differential neurotransmitter activity. *Sci Rep*. **8**, 4074 (2018).
127. F. J. García-Gómez, D. García-Solís, F. J. Luis-Simón, V. A. Marín-Oyaga, F. Carrillo, P. Mir, R. J. Vázquez-Albertino, [Elaboration of the SPM template for the standardization of SPECT images with 123I-Ioflupane]. *Rev Esp Med Nucl Imagen Mol*. **32**, 350–356 (2013).
128. S. Hesse, G.-A. Becker, M. Rullmann, A. Bresch, J. Luthardt, M. K. Hankir, F. Zientek, G. Reißig, M. Patt, K. Arelin, D. Lobsien, U. Müller, S. Baldofski, P. M. Meyer, M. Blüher, M. Fasshauer, W. K. Fenske, M. Stumvoll, A. Hilbert, Y.-S. Ding, O. Sabri, Central noradrenaline transporter availability in highly obese, non-depressed individuals. *Eur J Nucl Med Mol Imaging*. **44**, 1056–1064 (2017).
129. Y.-S. Ding, T. Singhal, B. Planeta-Wilson, J.-D. Gallezot, N. Nabulsi, D. Labaree, J. Ropchan, S. Henry, W. Williams, R. E. Carson, A. Neumeister, R. T. Malison, PET imaging of the effects of age and cocaine on the norepinephrine transporter in the human brain using (S,S)-[(11C)O-methylreboxetine and HRRT. *Synapse*. **64**, 30–38 (2010).
130. M. Nørgaard, V. Beliveau, M. Ganz, C. Svarer, L. H. Pinborg, S. H. Keller, P. S. Jensen, D. N. Greve, G. M. Knudsen, A high-resolution in vivo atlas of the human brain's benzodiazepine binding site of GABAA receptors. *Neuroimage*. **232**, 117878 (2021).
131. J. M. DuBois, O. G. Rousset, J. Rowley, M. Porras-Betancourt, A. J. Reader, A. Labbe, G. Massarweh, J.-P. Soucy, P. Rosa-Neto, E. Kobayashi, Characterization of age/sex and the regional distribution of mGluR5 availability in the healthy human brain measured by high-resolution [(11C)ABP688 PET. *Eur J Nucl Med Mol Imaging*. **43**, 152–162 (2016).
132. J. Y. Hansen, G. Shafiei, R. D. Markello, K. Smart, S. M. L. Cox, M. Nørgaard, V. Beliveau, Y. Wu, J.-D. Gallezot, É. Aumont, S. Servaes, S. G. Scala, J. M. DuBois, G. Wainstein, G. Bezgin, T. Funck, T. W. Schmitz, R. N. Spreng, M. Galovic, M. J. Koepp, J. S. Duncan, J. P.

- Coles, T. D. Fryer, F. I. Aigbirhio, C. J. McGinnity, A. Hammers, J.-P. Soucy, S. Baillet, S. Guimond, J. Hietala, M.-A. Bedard, M. Leyton, E. Kobayashi, P. Rosa-Neto, M. Ganz, G. M. Knudsen, N. Palomero-Gallagher, J. M. Shine, R. E. Carson, L. Tuominen, A. Dagher, B. Masic, Mapping neurotransmitter systems to the structural and functional organization of the human neocortex. *Nat Neurosci*, 1–13 (2022).
133. K. Smart, S. M. L. Cox, S. G. Scala, M. Tippler, N. Jaworska, M. Boivin, J. R. Séguin, C. Benkelfat, M. Leyton, Sex differences in [11C]ABP688 binding: a positron emission tomography study of mGlu5 receptors. *Eur J Nucl Med Mol Imaging*. **46**, 1179–1183 (2019).
  134. M. Galovic, A. Al-Diwani, U. Vivekananda, F. Torrealdea, K. Erlandsson, T. D. Fryer, Y. T. Hong, B. A. Thomas, C. J. McGinnity, E. Edmond, K. Sander, E. Årstad, I. Jelcic, F. I. Aigbirhio, A. M. Groves, K. Thielemans, B. Hutton, A. Hammers, J. S. Duncan, J. P. Coles, A. Barnes, C. J. Stagg, M. C. Walker, S. R. Irani, M. J. Koeppe, for the N. Investigators, In vivo NMDA receptor function in people with NMDA receptor antibody encephalitis (2021), doi:1v.
  135. A. T. Hillmer, I. Esterlis, J. D. Gallezot, F. Bois, M. Q. Zheng, N. Nabulsi, S. F. Lin, R. L. Papke, Y. Huang, O. Sabri, R. E. Carson, K. P. Cosgrove, Imaging of cerebral  $\alpha 4\beta 2^*$  nicotinic acetylcholine receptors with (-)-[(18F)]Flubatine PET: Implementation of bolus plus constant infusion and sensitivity to acetylcholine in human brain. *Neuroimage*. **141**, 71–80 (2016).
  136. M. Naganawa, N. Nabulsi, S. Henry, D. Matuskey, S.-F. Lin, L. Sliker, A. J. Schwarz, N. Kant, C. Jesudason, K. Ruley, A. Navarro, H. Gao, J. Ropchan, D. Labaree, R. E. Carson, Y. Huang, First-in-Human Assessment of 11C-LSN3172176, an M1 Muscarinic Acetylcholine Receptor PET Radiotracer. *Journal of Nuclear Medicine*. **62**, 553–560 (2021).
  137. M. Aghourian, C. Legault-Denis, J.-P. Soucy, P. Rosa-Neto, S. Gauthier, A. Kostikov, P. Gravel, M.-A. Bédard, Quantification of brain cholinergic denervation in Alzheimer’s disease using PET imaging with [18F]-FE0BV. *Mol Psychiatry*. **22**, 1531–1538 (2017).
  138. M.-A. Bedard, M. Aghourian, C. Legault-Denis, R. B. Postuma, J.-P. Soucy, J.-F. Gagnon, A. Pelletier, J. Montplaisir, Brain cholinergic alterations in idiopathic REM sleep behaviour disorder: a PET imaging study with 18F-FE0BV. *Sleep Med*. **58**, 35–41 (2019).
  139. T. Kantonen, T. Karjalainen, J. Isojärvi, P. Nuutila, J. Tuisku, J. Rinne, J. Hietala, V. Kaasinen, K. Kalliokoski, H. Scheinin, J. Hirvonen, A. Vehtari, L. Nummenmaa, Interindividual variability and lateralization of  $\mu$ -opioid receptors in the human brain. *NeuroImage*. **217**, 116922 (2020).
  140. J.-D. Gallezot, B. Planeta, N. Nabulsi, D. Palumbo, X. Li, J. Liu, C. Rowinski, K. Chidsey, D. Labaree, J. Ropchan, S.-F. Lin, A. Sawant-Basak, T. J. McCarthy, A. W. Schmidt, Y. Huang, R. E. Carson, Determination of receptor occupancy in the presence of mass dose: [11C]GSK189254 PET imaging of histamine H3 receptor occupancy by PF-03654746. *J Cereb Blood Flow Metab*. **37**, 1095–1107 (2017).
  141. M. D. Normandin, M.-Q. Zheng, K.-S. Lin, N. S. Mason, S.-F. Lin, J. Ropchan, D. Labaree, S. Henry, W. A. Williams, R. E. Carson, A. Neumeister, Y. Huang, Imaging the cannabinoid CB1 receptor in humans with [11C]OMAR: assessment of kinetic analysis methods, test-retest reproducibility, and gender differences. *J Cereb Blood Flow Metab*. **35**, 1313–1322 (2015).
  142. S. J. Finnema, N. B. Nabulsi, T. Eid, K. Detyniecki, S.-F. Lin, M.-K. Chen, R. Dhafer, D. Matuskey, E. Baum, D. Holden, D. D. Spencer, J. Mercier, J. Hannestad, Y. Huang, R. E. Carson, Imaging synaptic density in the living human brain. *Sci Transl Med*. **8**, 348ra96 (2016).
  143. E. H. Shen, C. C. Overly, A. R. Jones, The Allen Human Brain Atlas: Comprehensive gene expression mapping of the human brain. *Trends in Neurosciences*. **35**, 711–714 (2012).
  144. J. Dukart, S. Holiga, M. Rullmann, R. Lanzenberger, P. C. T. Hawkins, M. A. Mehta, S. Hesse, H. Barthel, O. Sabri, R. Jech, S. B. Eickhoff, JuSpace: A tool for spatial correlation analyses of magnetic resonance imaging data with nuclear imaging derived neurotransmitter maps. *Human Brain Mapping*. **42**, 555–566 (2021).
  145. R. D. Markello, J. Y. Hansen, Z.-Q. Liu, V. Bazinet, G. Shafiei, L. E. Suárez, N. Blostein, J. Seidlitz, S. Baillet, T. D. Satterthwaite, M. M. Chakravarty, A. Raznahan, B. Masic, neuromaps: structural and functional interpretation of brain maps. *Nat Methods*, 1–8 (2022).
  146. D. Wang, S. Liu, J. Warrell, H. Won, X. Shi, F. C. P. Navarro, D. Clarke, M. Gu, P. Emani, Y. T. Yang, M. Xu, M. J. Gandal, S. Lou, J. Zhang, J. J. Park, C. Yan, S. K. Rhie, K. Manakongtreecheep, H. Zhou, A. Nathan, M. Peters, E. Mattei, D. Fitzgerald, T. Brunetti, J. Moore, Y. Jiang, K. Girdhar, G. E. Hoffman, S. Kalayci, Z. H. Gümüş, G. E. Crawford, PsychENCODE Consortium, P. Roussos, S. Akbarian, A. E. Jaffe, K. P. White, Z. Weng, N. Sestan, D. H. Geschwind, J. A. Knowles, M. B. Gerstein, Comprehensive functional genomic resource and integrative model for the human brain. *Science*. **362**, eaat8464 (2018).
  147. B. B. Lake, R. Ai, G. E. Kaeser, N. S. Salathia, Y. C. Yung, R. Liu, A. Wildberg, D. Gao, H.-L. Fung, S. Chen, R. Vijayaraghavan, J. Wong, A. Chen, X. Sheng, F. Kaper, R. Shen, M. Ronaghi, J.-B. Fan, W. Wang, J. Chun, K. Zhang, Neuronal subtypes and diversity revealed by single-nucleus RNA sequencing of the human brain. *Science*. **352**, 1586–1590 (2016).
  148. D. Polioudakis, L. de la Torre-Ubieta, J. Langerman, A. G. Elkins, X. Shi, J. L. Stein, C. K. Vuong, S. Nichterwitz, M. Gevorgian, C. K. Opland, D. Lu, W. Connell, E. K. Ruzzo, J. K. Lowe, T. Hadzic, F. I. Hinz, S. Sabri, W. E. Lowry, M. B. Gerstein, K. Plath, D. H. Geschwind, A Single-Cell Transcriptomic Atlas of Human Neocortical Development during Mid-gestation. *Neuron*. **103**, 785-801.e8 (2019).
  149. J. A. Miller, S.-L. Ding, S. M. Sunkin, K. A. Smith, L. Ng, A. Szafer, A. Ebbert, Z. L. Riley, K. Aiona, J. M. Arnold, C. Bennet, D. Bertagnolli, K. Brouner, S. Butler, S. Caldejon, A. Carey, C. Cuhaciyan, R. A. Dalley, N. Dee, T. A. Dolbeare, B. A. C. Facer, D. Feng, T. P. Fliss, G. Gee, J. Goldy, L. Gourley, B. W. Gregor, G. Gu, R. E. Howard, J. M. Jochim, C. L. Kuan, C. Lau, C.-K. Lee, F. Lee, T. A. Lemon, P. Lesnar, B. McMurray, N. Mastan, N. F. Mosqueda, T. Nalua-Cecchini, N.-K. Ngo, J. Nyhus, A. Oldre, E. Olson, J. Parente, P. D. Parker, S. E. Parry, A. S. Player, M. Pletikos, M. Reding, J. J. Royall, K. Roll, D. Sandman, M. Sarreal, S. Shapouri, N. V. Shapovalova, E. H. Shen, N. Sjoquist, C. R. Slaughterbeck, M. Smith, A. J. Sodr, D. Williams, L. Zöllei, B. Fischl, M. B. Gerstein, D. H. Geschwind, I. A. Glass, M. J. Hawrylycz, R. F. Hevner, H. Huang, A. R. Jones, J. A. Knowles, P. Levitt, J. W. Phillips, N. Sestan, P. Wohnoutka, C. Dang, A. Bernard, J. G. Hohmann, E. S. Lein, Transcriptional Landscape of the Prenatal Human Brain. *Nature*. **508**, 199–206 (2014).
  150. J. Piñero, J. M. Ramirez-Anguita, J. Saüch-Pitarch, F. Ronzano, E. Centeno, F. Sanz, L. I. Furlong, The DisGeNET knowledge platform for disease genomics: 2019 update. *Nucleic Acids Research*. **48**, D845–D855 (2020).
  151. M. Ashburner, C. A. Ball, J. A. Blake, D. Botstein, H. Butler, J. M. Cherry, A. P. Davis, K. Dolinski, S. S. Dwight, J. T. Eppig, M. A. Harris, D. P. Hill, L. Issel-Tarver, A. Kasarskis, S. Lewis, J. C. Matese, J. E. Richardson, M. Ringwald, G. M. Rubin, G. Sherlock, Gene Ontology: tool for the unification of biology. *Nat Genet*. **25**, 25–29 (2000).
  152. The Gene Ontology Consortium, S. Carbon, E. Douglass, B. M. Good, D. R. Unni, N. L. Harris, C. J. Mungall, S. Basu, R. L. Chisholm, R. J. Dodson, E. Hartline, P. Fey, P. D. Thomas, L.-P. Albou, D. Ebert, M. J. Kesling, H. Mi, A. Muruganujan, X. Huang, T. Mushayahama, S. A. LaBonte, D. A. Siegele, G. Antonazzo, H. Attrill, N. H. Brown, P. Garapati, S. J. Marygold, V. Trovisco, G. dos Santos, K. Falls, C. Tabone, P. Zhou, J. L. Goodman, V. B. Strelets, J. Thurmond, P. Garmiri, R. Ishtiaq, M. Rodríguez-López, M. L. Acencio, M. Kuiper, A. Lægreid, C. Logie, R. C. Lovering, B. Kramarz, S. C. C. Saverimuttu, S. M. Pinheiro, H. Gunn, R. Su, K. E. Thurlow, M. Chibucos, M. Glioglio, S. Nadendla, J. Munro, R. Jackson, M. J. Duesbury, N. Del-Toro, B. H. M. Meldal, K. Paneerselvam, L. Perfetto, P. Porras, S. Orchard, A. Shrivastava, H.-Y. Chang, R. D. Finn, A. L. Mitchell, N. D. Rawlings, L. Richardson, A. Sangrador-Vegas, J. A. Blake, K. R. Christie, M. E. Dolan, H. J. Drabkin, D. P. Hill, L. Ni, D. M. Sitnikov, M. A. Harris, S. G. Oliver, K. Rutherford, V. Wood, J. Hayles, J. Bähler, E. R. Bolton, J. L. De Pons, M. R. Dwinell, G. T. Hayman, M. L. Kaldunski, A. E. Kwitek, S. J. F. Laulederkind, C. Plasterer,

- M. A. Tutaj, M. Vedi, S.-J. Wang, P. D'Eustachio, L. Matthews, J. P. Balhoff, S. A. Aleksander, M. J. Alexander, J. M. Cherry, S. R. Engel, F. Gondwe, K. Karra, S. R. Miyasato, R. S. Nash, M. Simison, M. S. Skrzypek, S. Weng, E. D. Wong, M. Feuermann, P. Gaudet, A. Morgat, E. Bakker, T. Z. Berardini, L. Reiser, S. Subramaniam, E. Huala, C. N. Arighi, A. Auchincloss, K. Axelsen, G. Argoud-Puy, A. Bateman, M.-C. Blatter, E. Boutet, E. Bowler, L. Breuza, A. Bridge, R. Britto, H. Bye-A-Jee, C. C. Casas, E. Coudert, P. Denny, A. Estreicher, M. L. Famiglietti, G. Georghiou, A. Gos, N. Gruaz-Gumowski, E. Hatton-Ellis, C. Hulo, A. Ignatchenko, F. Jungo, K. Laiho, P. Le Mercier, D. Lieberherr, A. Lock, Y. Lussi, A. MacDougall, M. Magrane, M. J. Martin, P. Masson, D. A. Natale, N. Hyka-Nouspikel, S. Orchard, I. Pedruzzi, L. Pourcel, S. Poux, S. Pundir, C. Rivoire, E. Speretta, S. Sundaram, N. Tyagi, K. Warner, R. Zaru, C. H. Wu, A. D. Diehl, J. N. Chan, C. Grove, R. Y. N. Lee, H.-M. Muller, D. Raciti, K. Van Auken, P. W. Sternberg, M. Berriman, M. Paulini, K. Howe, S. Gao, A. Wright, L. Stein, D. G. Howe, S. Toro, M. Westerfield, P. Jaiswal, L. Cooper, J. Elser, The Gene Ontology resource: enriching a Gold mine. *Nucleic Acids Research*. **49**, D325–D334 (2021).
153. B. D. Fulcher, A. Arnatkeviciute, A. Fornito, Overcoming false-positive gene-category enrichment in the analysis of spatially resolved transcriptomic brain atlas data. *Nat Commun*. **12**, 2669 (2021).
  154. M. J. M. F. Reijnders, R. M. Waterhouse, Summary Visualizations of Gene Ontology Terms With GO-Figure! *Frontiers in Bioinformatics*. **1** (2021), doi:10.3389/fbinf.2021.638255.
  155. S. Darmanis, S. A. Sloan, Y. Zhang, M. Enge, C. Caneda, L. M. Shuer, M. G. Hayden Gephart, B. A. Barres, S. R. Quake, A survey of human brain transcriptome diversity at the single cell level. *Proc Natl Acad Sci USA*. **112**, 7285–7290 (2015).
  156. S. Grote, K. Prüfer, J. Kelso, M. Dannemann, ABAEnrichment: an R package to test for gene set expression enrichment in the adult and developing human brain. *Bioinformatics*. **32**, 3201–3203 (2016).
  157. X. Jiao, B. T. Sherman, D. W. Huang, R. Stephens, M. W. Baseler, H. C. Lane, R. A. Lempicki, DAVID-WS: a stateful web service to facilitate gene/protein list analysis. *Bioinformatics*. **28**, 1805–1806 (2012).
  158. T. Salo, K. L. Bottenhorn, T. E. Nichols, M. C. Riedel, M. T. Sutherland, T. Yarkoni, A. R. Laird, NiMARE: a neuroimaging meta-analysis research environment. *F1000Research*. **7** (2018), doi:10.7490/f1000research.1115905.1.
  159. C. Feng, S. B. Eickhoff, T. Li, L. Wang, B. Becker, J. A. Camilleri, S. Héту, Y. Luo, Common brain networks underlying human social interactions: evidence from large-scale neuroimaging meta-analysis. *Neuroscience & Biobehavioral Reviews* (2021), doi:10.1016/j.neubiorev.2021.03.025.
  160. M. Schurz, J. Radua, M. G. Tholen, L. Maliske, D. S. Margulies, R. B. Mars, J. Sallet, P. Kanske, Toward a hierarchical model of social cognition: A neuroimaging meta-analysis and integrative review of empathy and theory of mind. *Psychol Bull*. **147**, 293–327 (2021).
  161. L. Ficco, L. Mancuso, J. Manuello, A. Teneggi, D. Liloia, S. Duca, T. Costa, G. Z. Kovacs, F. Cauda, Disentangling predictive processing in the brain: a meta-analytic study in favour of a predictive network. *Sci Rep*. **11**, 16258 (2021).
  162. D. Bzdok, R. Langner, L. Schilbach, O. Jakobs, C. Roski, S. Caspers, A. R. Laird, P. T. Fox, K. Zilles, S. B. Eickhoff, Characterization of the temporo-parietal junction by combining data-driven parcellation, complementary connectivity analyses, and functional decoding. *NeuroImage*. **81**, 381–392 (2013).



versions. *A/V* indicates whether participants in the experiments had contact per audio and/or video during fMRI scanning (live transmission in the case of hyperscanning experiments). *Method* refers to the general type of analysis method: *Seed*: seed-to-voxel (whole-brain), foci within the “target-brain” were included. *Wb-ICA*: ICA on whole-brain level, foci were peak coordinates of reported independent components. *ts-ICA*: only independent components sensitive to the applied task were analyzed. *wb-VW*: voxel-wise analysis on whole-brain level. *ts-VW*: voxel-wise analysis on voxels sensitive to the applied task. *wb-Atlas*: a whole-brain atlas was applied, foci correspond to center-of-masses of reported parcels. *Lag* refers to whether time-series of interacting subjects were analyzed with time lag. If possible, we restricted the included foci to those from “zero lag” analyses. *Band* indicates whether a band pass or high pass filter was applied. *Contributions* refers to the relative contribution of each study to each cluster in the main fMRI ALE using the conservative voxel-level threshold ( $p < .001$ ) or the lenient threshold ( $p < .01$ ), and the combined fMRI and fNIRS ALE using the conservative thresholding. The ALE method incorporates nonlinear procedures causing the percentages to not add up to 100%.

Abbreviations: ALE = activation likelihood estimation, hyper(3) = hyperscanning (in one case between 3 subjects), pseudo = pseudo-hyperscanning, A = audio, V = video, INS = interpersonal neural synchronization, JA = joint attention, JAct = joint action, y = years, MNI = Montreal Neurological Institute space, TAL = Talairach space, wb = whole-brain, ts = task-specific, ICA = independent component analysis, VW = voxel-wise, TR = repetition time, rTPJ = right temporoparietal junction, rSTG = right superior temporal gyrus, rIns = right insula, r/ISFG = right/left superior frontal gyrus, lMFG = left medial frontal gyrus.



**Table S2: fNIRS experiments included for meta-analysis**

See the separate supplementary file for the original table in Excel format. Data from 54 hyperscanning fNIRS publications (3, 5, 60–111) and 3 pseudohyperscanning fNIRS publications (112–114) were included in the analyses. *Publications* refers to the single publications (n = 57) that were included in meta-analyses. *Experiment ID* refers to the aggregated experiments when considering separately analyzed subgroups and reanalyses of existing data (n = 69 experiments). *Setting* indicates how participants were seated during the experiment. When subjects were seated side-by-side or face-to-face, they nevertheless often were separated by a portable wall or similar. *Length* refers to the wavelength of the light emitted by the fNIRS device. *Band* indicates how band pass filtering was applied. *Coverage* refers to the general placement of fNIRS probe arrays on the head. Multiple entries indicate multiple separated probe arrays. *Array* refers to the desing of the probe array, “custom” indicates a non-square format. *Sources* lists the source of fNIRS coordinates: AV: AtlasViewer (115), Dig: 3D digitizer, VR: virtual registration (2), MRI: anatomical MRI-based registration, or from another publication. For “Dig” and “MRI”, numbers in brackets following the abbreviations indicate the number of subjects in the sample from which coordinates were estimated. For “AV”, numbers in brackets show reconstruction quality coded as: 3 = all optodes have assigned 10-10 positions, 2 = at least two 10-10 positions for each optode array, 1 = one 10-10 positions and information about the array alignment. *Contributions* refers to the relative contribution of each study to each cluster in the ALE analysis on the combined fMRI and fNIRS data. Of note, the ALE method incorporates nonlinear procedures causing the percentages to not add up to 100%.

Abbreviations: ALE = activation likelihood estimation, hyper = hyperscanning, pseudo = pseudo-hyperscanning, INS = interpersonal neural synchronization, y = years, rTPJ = right temporoparietal junction, r/ISFG = right/left superior frontal gyrus, lMFG = left medial frontal gyrus.

Target system	Target function	Atlases		Subjects (N)	Mean age (y)	Source
		Name	Tracer			
<b>Serotonin</b>	serotonin receptor 1a	5HT1a	(11C)WAY100635	35	26.3	Savli et al., 2012 (118)
	serotonin receptor 1b	5HT1b	(11C)P943	65, 23	33.7, 28.7	Gallezot et al., 2010; Savli et al., 2012 (118, 119)
	serotonin receptor 2a	5HT2a	(11C)Cimbi-36	29	22.6	Beliveau et al., 2017 (120)
	serotonin receptor 4	5HT4	(11C)SB207145	59	25.9	Beliveau et al., 2017 (120)
	serotonin receptor 6	5HT6	(11C)GSK215083	30	36.6	Radhakrishnan et al., 2018 (121)
	serotonin transporter	5HTT	(11C)DASB	100, 18	25.1, 30.5	Beliveau et al., 2017; Savli et al., 2012 (118, 120)
<b>Dopamine</b>	dopamine synthesis	FDOPA	(18F)fluorodopa	12	n.a.	García-Gómez et al., 2018 (122)
	dopamine receptor 1	D1	(11C)SCH23390	13	33	Kaller et al., 2017 (123)
	dopamine receptor 2	D2	(11C)FLB-457	37, 55	48.4, 32.5	Sandiego et al., 2015; Smith et al., 2019 (124, 125)
	dopamine transporter	DAT	(123I)FP-CIT	174, 30	61, n.a.	Dukart et al., 2018; García-Gómez et al., 2013 (126, 127)
<b>Noradrenaline</b>	noradrenaline transporter	NET	(11C)O-MRB	77, 10	33.4, 33.3	Hesse et al., 2017; Ding et al., 2010 (128, 129)
<b>GABA</b>	GABA receptor A	GABAa	(11C)flumazenil	6, 16	n.a., 26.6	Dukart et al., 2018; Nørgaard et al., 2021 (126, 130)
<b>Glutamate</b>	metabotropic receptor 5	mGluR5	(11C)ABP688	73, 22, 28	19.9, 67.9, 33.1	DuBois et al., 2016; Hansen et al., 2022; Smart et al., 2019 (131–133)
	NMDA receptor	NMDA	(18F)GE-179	29	41	Galovic et al., 2021 (134)
<b>Acetylcholine</b>	$\alpha$ 4 $\beta$ 2 nicotinic receptor	a4b2	(F18)flubatine	30	33.5	Hillmer et al., 2016 (135)
	muscarinic receptor 1	M1	(11C)LSN3172176	24	40.5	Naganawa et al., 2021 (136)
	vesicular Ach transporter	VachT	(18F)FEOBV	4, 18, 5	37, 66.8, 68.3	Aghourian et al., 2017; Bedard et al., 2019; Hansen et al., 2022 (132, 137, 138)
<b>Endorphins</b>	$\mu$ receptor	MU	(11C)carfentanil	204, 39	32.3, n.a.	Hansen et al., 2022; Kantonen et al., 2020 (132, 139)
<b>Histamine</b>	histamine receptor 3	H3	(11C)GSK189254	8	31.7	Gallezot et al., 2017 (140)
<b>Cannabinoid</b>	cannabinoid receptor 1	CB1	(11C)OMAR	77	30	Normandin et al., 2015 (141)
<b>Synaptic density</b>	vesicle glycoprotein 2A	SV2a	(11C)UCB-J	10	36	Finnema et al., 2016 (142)
<b>Oxytocin</b>	oxytocin	OXT				
	oxytocin receptor	OXTR	mRNA expression	up to 6	24 – 57	Shen et al., 2012; Markello et al., 2021 (143, 20)
	oxytocin release	CD38				

**Table S3: Sources of neurotransmitter maps**

All nuclear imaging *in vivo* neurotransmitter atlases are drawn from JuSpace or neuromaps (144, 145), mRNA expression atlases are derived from the Allen brain atlas (143) using the abagen toolbox with default settings (20). Sources listed in the table are the original sources of the atlases. For spatial correlation analyses, the atlases were parcellated, Z-standardized, and the weighted mean was calculated if multiple atlases using the same tracer were available.

Abbreviations: y = years.



Analysis	Dataset	Sources	N (categories)		N (genes)
			all	included	
<b>Neuronal cell types</b>	<i>PsychENCODE</i> cell markers (transcripts per kilobase counts)	Wang et al., 2018; Darmanis et al., 2015; Lake et al., 2016 ( <i>146, 147, 155</i> )	24	24	2 – 83
<b>Developmental regional enrichment</b>	<i>BrainSpan</i> expression data (expression > 0.9 <sup>th</sup> quantile, annotated to ≤ 20% of categories)	Miller et al., 2014; Grote et al., 2016 ( <i>149, 156</i> )	80	80	5 – 224
<b>Psychiatric disorders</b>	<i>DisGeNET</i> disease markers (manually curated dataset)	Piñero et al., 2020; Jiao et al., 2012 ( <i>150, 157</i> )	332	150	5 – 697
<b>Biological functions</b>	<i>Gene Ontology</i> biological processes (annotations propagated through the hierarchy, retrieved March 2022)	Ashburner et al., 2000; The Gene Ontology Consortium et al., 2021; Jiao et al., 2012 ( <i>151, 152, 157</i> )	15,039	6,947	5 – 200

**Table S4: Gene category enrichment datasets**

All datasets, except for *BrainSpan* data, were available as genetic “markers”. Gene-wise expression values in the *BrainSpan* dataset were thresholded according to the above settings to identify markers for each of 80 categories. *DisGeNET* and *BrainSpan* categories were thresholded to contain at least 5 genes, *Gene Ontology* categories were thresholded to contain between 5 and 200 genes (*153*).

URLs: *PsychENCODE*: <http://resource.psychencode.org/>, *BrainSpan*: <https://www.brainspan.org/>, *ABAEnrichment*: <https://bioconductor.org/packages/ABAEnrichment/>, *DisGeNET*: <https://www.disgenet.org/>, *Gene Ontology*: <http://geneontology.org/>, *DAVID*: <https://david.ncifcrf.gov/>.

Analysis	Cluster	X	Y	Z	Estimate	Volume	AAL regions
ALE ( $p < .001$ )	rTPJ	62	-48	16	0.019	2856	58.26% Temporal Mid R; 36.41% Temporal Sup R
	rSTG	50	-20	-6	0.017	752	82.98% Temporal Sup R; 15.96% Temporal Mid R
ALE ( $p < .01$ )	rTPJ	62	-48	16	0.041	7856	47.45% Temporal Mid R; 25.25% Temporal Sup R; 12.02% SupraMarginal R; 8.66% Temporal Inf R
	rSTG	50	-20	-6	0.025	3368	88.84% Temporal Sup R; 8.55% Temporal Mid R
	rIns	38	20	-2	0.021	3296	52.43% Insula R; 18.69% Frontal Inf Orb 2 R; 13.11% Frontal Inf Tri R; 10.19% Frontal Inf Oper R; 5.58% no label
ALE (no pseudo)	rTPJ	62	-48	16	0.016	2152	63.94% Temporal Mid R; 32.71% Temporal Sup R
	rTPJ	62	-48	16	0.019	3440	63.95% Temporal Mid R; 34.42% Temporal Sup R
ALE & fNIRS (all fNIRS)	ISFG	-26	66	8	0.021	2248	64.41% Frontal Sup 2 L; 20.28% no_label; 10.32% Frontal Sup Medial L
	rSFG	28	54	32	0.020	2040	47.84% Frontal Sup 2 R; 23.92% Frontal Sup Medial R; 23.14% Frontal Mid 2 R; 5.10% no label
	rMFG	36	38	42	0.021	784	100.00% Frontal Mid 2 R
	rTPJ	60	-48	16	0.022	3368	63.42% Temporal Mid R; 34.92% Temporal Sup R
ALE & fNIRS (restricted fNIRS)	ISFG	-26	66	8	0.021	2064	64.34% Frontal Sup 2 L; 19.38% no_label; 10.85% Frontal Sup Medial L
	rMFG/ rSFG	28	54	32	0.021	824	51.46% Frontal Mid 2 R; 42.72% Frontal Sup 2 R; 5.83% no label
	rMFG	36	38	42	0.021	792	100.00% Frontal Mid 2 R
MACM	rTPJ	60	-48	16	0.085	28576	39.39% Temporal Mid R; 18.20% Temporal Sup R; 11.62% SupraMarginal R; 9.55% Temporal Inf R; 7.84% Fusiform R; 6.63% Occipital Inf R
	ITPJ	-58	-42	22	0.066	26280	35.43% Temporal Mid L; 17.08% SupraMarginal L; 15.68% Temporal Sup L; 8.80% Fusiform L; 5.02% Temporal Inf L
	IPFCIns	-32	20	2	0.068	22456	24.90% Frontal Inf Tri L; 21.66% Insula L; 18.28% Precentral L; 15.57% Frontal Inf Oper L; 12.22% Frontal Inf Orb 2 L
	rPFCIns	36	20	-2	0.069	19760	22.43% Frontal Inf Oper R; 19.64% Insula R; 19.51% Precentral R; 18.70% Frontal Inf Tri R; 6.80% Frontal Inf Orb 2 R; 5.71% no label
	SMA	-4	12	48	0.070	11416	32.94% Supp_Motor_Area L; 24.04% Supp_Motor_Area R; 20.88% Cingulate Mid R; 9.39% Cingulate Mid L; 8.20% Frontal Sup Medial L
	lTh	-10	-18	4	0.066	5672	54.72% Thalamus L; 16.50% no_label; 14.10% Hippocampus L; 10.86% Amygdala L
	lIPL	-34	-50	46	0.061	5448	64.32% Parietal Inf L; 27.02% Parietal Sup L; 5.87% Precuneus L
	rTh	10	-16	6	0.061	4664	49.40% Thalamus R; 24.19% no_label; 17.67% Pallidum R; 6.69% Caudate R
	rIPL	32	-58	48	0.061	2872	37.33% Parietal Inf R; 31.20% Angular R; 30.08% Parietal Sup R
	lPrec	-2	-56	34	0.063	1520	50.53% Precuneus L; 31.58% Cingulate Post L; 15.26% Precuneus R

**Table S5: Information on clusters resulting from ALE and MACM analyses**

Data derived from ALE maps after application of a cluster-level threshold at familywise error-corrected  $p < .05$ . AtlasReader (*116*) was used to estimate peak MNI coordinates ( $X$ ,  $Y$ ,  $Z$ ), the average ALE value (*Estimate*), the cluster volume in mm (*Volume*), and coverage of anatomical regions relative to the cluster according to the AAL atlas (*117*). Abbreviations: ALE = activation likelihood estimation, pseudo = pseudo-hyperscanning, fNIRS = functional near-infrared spectroscopy, MACM = meta-analytic connectivity modelling, AAL = automated anatomic labelling atlas, r/ITPJ = right/left temporoparietal junction, rSTG = right superior temporal gyrus, rIns = right insula, r/IPFCIns = right/left prefrontal cortex-insula, SMA = supplementary motor area, r/lTh = left thalamus, r/lIPL = left inferior parietal lobule, lPrec = left precuneus.

## All included fNIRS experiments

Parcel	N (ch)				N (exp)				N (sub)				Ratio * N (sub-all)				Ratio * N (exp-all)				AAL region
	all	sync	ratio	p	p (M)	p (%)	all	sync	all	sync	value	p	p (M)	p (%)	value	p	p (M)	p (%)			
RH Vis 3	19	4	0.21	0.169	0.320	0.024	18	4	1339	592	281.89	0.014	0.052	0.481	3.79	0.075	0.236	0.056	52.97%	Temporal Inf R	
LH Default PFC 2	56	11	0.20	0.057	0.092	0.295	44	7	2205	268	433.13	0.018	0.116	0.181	8.64	0.007	0.091	0.303	37.52%	Frontal Inf Tri L	
LH DorsAttn Post 6	5	3	0.60	0.013	0.058	0.480	4	3	154	118	92.40	0.047	0.149	0.114	2.40	0.012	0.102	0.288	57.52%	Parietal Sup L	
RH SalVentAttn TempOccPar 1	35	5	0.14	0.437	0.556	<0.001	31	4	2145	176	306.43	0.048	0.280	0.079	4.43	0.188	0.484	0.001	53.36%	Temporal Sup R	
RH_Default_PFCdPFCm_1	38	6	0.16	0.264	0.477	0.011	38	6	1844	280	291.16	0.076	0.317	0.027	6.00	0.044	0.293	0.066	29.90%	Frontal Med Orb R	
LH SalVentAttn FrOperIns 2	4	1	0.25	0.396	0.999	0.004	3	1	242	18	60.50	0.111	0.999	0.107	0.75	0.395	0.999	0.004	64.87%	Insula L	
RH_Default_PFCdPFCm_2	114	17	0.15	0.151	0.295	0.006	46	13	2818	1002	420.23	0.124	0.339	0.002	6.86	0.278	0.555	<0.001	41.38%	Frontal Sup Medial R	
LH Default Temp 2	6	1	0.17	0.544	0.768	<0.001	6	1	444	48	74.00	0.142	0.612	<0.001	1.00	0.457	0.733	<0.001	88.43%	Temporal Mid L	
RH Cont PFC1 3	87	12	0.14	0.319	0.349	0.034	47	11	2756	912	380.14	0.146	0.215	0.066	6.48	0.259	0.388	0.020	65.37%	Frontal Mid 2 R	
LH Default PFC 4	113	21	0.19	0.015	0.029	0.691	43	12	2098	506	389.89	0.205	0.228	0.007	7.99	0.102	0.132	0.086	55.26%	Frontal Sup 2 L	
RH Cont PFC1 2	61	7	0.11	0.599	0.652	<0.001	42	5	2545	206	292.05	0.222	0.377	0.014	4.82	0.413	0.557	0.001	52.90%	Frontal Mid 2 R	
LH Cont Par 1	9	3	0.33	0.076	0.196	0.178	6	2	212	66	70.67	0.258	0.270	0.024	2.00	0.186	0.225	0.124	71.29%	Parietal Inf L	
RH Cont Par 2	63	9	0.14	0.311	0.397	0.002	32	8	2015	503	287.86	0.263	0.476	0.001	4.57	0.459	0.650	<0.001	59.87%	Angular R	
RH Limbic OFC 1	4	1	0.25	0.376	0.404	0.056	4	1	146	18	36.50	0.263	0.248	0.080	1.00	0.341	0.241	0.104	28.24%	Rectus R	
RH Default Temp 3	10	1	0.10	0.713	0.761	<0.001	9	1	734	180	73.40	0.278	0.456	0.028	0.90	0.657	0.660	0.004	48.97%	Temporal Mid R	
RH_Default_Temp_2	4	1	0.25	0.378	0.504	<0.001	4	1	131	88	32.75	0.297	0.430	<0.001	1.00	0.343	0.470	<0.001	43.90%	Temporal Pole Sup R	
LH Default PFC 6	9	2	0.22	0.300	0.287	0.063	8	2	308	66	68.44	0.301	0.304	0.003	1.78	0.283	0.232	0.066	73.85%	Frontal Mid 2 L	
RH Default PFCv 2	23	5	0.22	0.117	0.038	0.590	14	3	631	79	137.17	0.307	0.135	0.107	3.04	0.241	0.103	0.188	49.04%	Frontal Inf Tri R	
LH Cont PFC1 1	63	8	0.13	0.444	0.401	0.021	45	7	2075	226	263.49	0.337	0.384	0.007	5.71	0.232	0.325	0.030	54.13%	Frontal Inf Tri L	
LH SalVentAttn ParOper 1	18	5	0.28	0.053	0.183	0.108	12	4	399	148	110.83	0.341	0.609	0.001	3.33	0.107	0.385	0.008	54.68%	SupraMarginal L	
RH Default Par 1	48	5	0.10	0.688	0.718	<0.001	33	4	2063	120	214.90	0.384	0.481	0.005	3.44	0.602	0.702	<0.001	39.79%	Angular R	
RH_DorsAttn_PrCv_1	22	5	0.23	0.096	0.400	<0.001	16	5	513	186	116.59	0.389	0.499	<0.001	3.64	0.135	0.417	<0.001	44.81%	Frontal Inf Oper R	
LH DorsAttn Post 2	18	3	0.17	0.339	0.369	0.013	14	3	531	102	88.50	0.427	0.517	<0.001	2.33	0.329	0.379	0.007	29.06%	Parietal Inf L	
LH SalVentAttn PFC1 1	76	10	0.13	0.384	0.551	<0.001	43	10	2042	436	268.68	0.440	0.674	<0.001	5.66	0.338	0.607	<0.001	58.16%	Frontal Mid 2 L	
LH Default Par 1	16	2	0.13	0.579	0.296	0.042	15	2	647	197	80.88	0.455	0.288	0.017	1.88	0.401	0.262	0.046	69.08%	Temporal Mid L	
LH Vis 7	6	1	0.17	0.503	0.666	<0.001	6	1	163	48	27.17	0.478	0.661	<0.001	1.00	0.419	0.661	<0.001	42.60%	Temporal Mid L	
RH Vis 7	31	4	0.13	0.496	0.587	0.004	20	3	1040	119	134.19	0.493	0.537	0.013	2.58	0.514	0.646	0.001	74.44%	Occipital Mid R	
RH SomMot 4	23	3	0.13	0.494	0.754	<0.001	17	3	741	65	96.65	0.514	0.702	<0.001	2.22	0.477	0.714	<0.001	62.76%	Postcentral R	
LH DorsAttn Post 3	8	1	0.13	0.641	0.549	0.007	4	1	138	18	17.25	0.641	0.608	<0.001	0.50	0.641	0.602	<0.001	48.76%	Parietal Sup L	
LH SomMot 5	19	4	0.21	0.165	0.223	0.055	6	4	246	166	51.79	0.722	0.705	<0.001	1.26	0.69	0.654	<0.001	55.07%	Postcentral L	
RH Default PFCdPFCm 3	22	2	0.09	0.736	0.813	<0.001	13	1	584	32	53.09	0.759	0.743	<0.001	1.18	0.736	0.780	<0.001	61.82%	Frontal Sup 2 R	
RH SomMot 1	31	2	0.06	0.895	0.926	<0.001	25	2	1333	105	86.00	0.766	0.807	<0.001	1.61	0.793	0.882	<0.001	61.86%	Temporal Sup R	
RH DorsAttn Post 3	19	1	0.05	0.915	0.824	<0.001	17	1	826	32	43.47	0.777	0.744	<0.001	0.89	0.774	0.808	<0.001	42.84%	Parietal Inf R	
LH SomMot 4	25	2	0.08	0.812	0.930	<0.001	18	2	657	44	52.56	0.832	0.928	<0.001	1.44	0.802	0.927	<0.001	73.10%	Postcentral L	
RH_Cont_Par_1	49	3	0.06	0.940	0.887	<0.001	30	3	1845	550	112.96	0.855	0.808	<0.001	1.84	0.936	0.875	<0.001	59.43%	SupraMarginal_R	

RH DorsAttn Post 5	25	2	0.08	0.831	0.806	<0.001	15	2	606	49	48.48	0.859	0.804	<0.001	1.20	0.831	0.803	<0.001	46.68% Precuneus R
RH DorsAttn Post 2	17	1	0.06	0.894	0.887	<0.001	12	1	534	44	31.41	0.860	0.827	<0.001	0.71	0.891	0.853	0.001	85.28% Postcentral R
RH SalVentAttn TempOccPar 2	50	4	0.08	0.835	0.780	<0.001	29	3	1295	112	103.60	0.884	0.822	<0.001	2.32	0.838	0.799	<0.001	68.61% SupraMarginal R
LH Default Par 2	47	10	0.21	0.037	0.020	0.741	13	6	447	132	95.11	0.890	0.878	<0.001	2.77	0.743	0.685	<0.001	36.97% Angular L
RH DorsAttn Post 1	47	3	0.06	0.918	0.553	0.005	26	2	1365	152	87.13	0.905	0.432	0.018	1.66	0.916	0.675	<0.001	64.75% Temporal Mid R
RH DorsAttn Post 4	40	3	0.08	0.871	0.945	<0.001	20	3	978	191	73.35	0.915	0.957	<0.001	1.50	0.907	0.956	<0.001	56.13% Parietal Sup R
RH Cont PFC1 1	169	19	0.11	0.617	0.491	<0.001	43	10	2052	398	230.70	0.959	0.813	<0.001	4.83	0.913	0.768	<0.001	44.35% Frontal Sup 2 R
LH SomMot 1	28	1	0.04	0.968	0.931	<0.001	17	1	747	18	26.68	0.967	0.919	<0.001	0.61	0.968	0.929	<0.001	77.45% Temporal Sup L
RH SomMot 6	50	3	0.06	0.940	0.955	<0.001	21	2	1022	50	61.32	0.977	0.951	<0.001	1.26	0.987	0.955	<0.001	54.07% Precentral R
LH_Default_PFC_5	159	9	0.06	0.999	0.993	<0.001	47	7	2226	350	126.00	0.999	0.997	<0.001	2.66	0.999	0.996	<0.001	51.49% Frontal Sup Medial L

## fNIRS experiments restricted to specific INS &gt; rest, control, or randomization

Parcel	N (ch)				N (exp)		N (sub)		Ratio * N (sub-all)		Ratio * N (exp-all)		AAL region
	all	sync	ratio	p	all	sync	all	sync	value	p	value	p	
LH Default PFC 4	100	19	0.19	0.008	33	6	1514	280	275.27	0.008	7.03	0.051	55.26% Frontal Sup 2 L
RH Cont PFC1 3	76	11	0.14	0.205	4	3	154	118	92.40	0.021	5.93	0.130	65.37% Frontal Mid 2 R
RH_Default_PFCdPFCm_1	33	6	0.18	0.147	37	11	1704	484	323.76	0.027	6.00	0.015	29.90% Frontal Med Orb R
RH_Default_PFCdPFCm_2	95	12	0.13	0.357	37	5	1613	216	245.46	0.052	5.05	0.384	41.38% Frontal Sup Medial R
LH Default PFC 2	46	7	0.15	0.246	41	10	1946	396	281.66	0.063	5.63	0.048	37.52% Frontal Inf Tri L
LH SalVentAttn PFC1 1	60	8	0.13	0.315	15	3	637	76	127.40	0.088	4.40	0.306	58.16% Frontal Mid 2 L
RH Cont PFC1 2	50	6	0.12	0.465	5	2	182	66	78.00	0.090	4.44	0.210	52.90% Frontal Mid 2 R
LH Cont PFC1 1	46	6	0.13	0.386	23	3	1059	154	162.92	0.112	4.57	0.176	54.13% Frontal Inf Tri L
RH Cont PFC1 1	154	17	0.11	0.544	35	5	1569	174	204.65	0.144	4.19	0.843	44.35% Frontal Sup 2 R
RH SalVentAttn TempOccPar 1	26	4	0.15	0.313	37	5	1727	206	207.24	0.153	3.54	0.143	53.36% Temporal Sup R
RH Cont Par 2	51	7	0.14	0.343	4	1	176	48	44.00	0.156	3.57	0.475	59.87% Angular R
RH Vis 3	15	3	0.20	0.225	2	1	62	18	31.00	0.161	3.00	0.081	52.97% Temporal Inf R
RH Default Par 1	35	3	0.09	0.783	40	10	1972	428	249.09	0.207	2.23	0.634	39.79% Angular R
RH Default PFCv 2	16	5	0.31	0.039	33	8	1566	382	208.80	0.217	3.13	0.100	49.04% Frontal Inf Tri R
LH DorsAttn Post 6	5	3	0.60	0.007	4	1	146	18	36.50	0.224	2.40	0.006	57.52% Parietal Sup L
RH Vis 7	27	3	0.11	0.579	10	3	311	79	97.19	0.238	1.78	0.587	74.44% Occipital Mid R
RH DorsAttn Post 1	36	3	0.08	0.769	11	3	365	134	73.00	0.369	1.75	0.768	64.75% Temporal Mid R
LH_Default_PFC_5	135	6	0.04	0.997	9	2	311	90	51.83	0.389	1.69	0.999	51.49% Frontal Sup Medial L
LH Default Par 2	35	9	0.26	0.013	6	1	163	48	27.17	0.456	2.57	0.502	36.97% Angular L
LH Cont Par 1	7	3	0.43	0.035	26	6	1131	361	155.24	0.461	2.14	0.035	71.29% Parietal Inf L
RH_DorsAttn_PrCv_1	15	3	0.20	0.231	12	2	415	43	51.88	0.541	2.20	0.227	44.81% Frontal Inf Oper R
RH SomMot 6	46	3	0.07	0.901	16	2	760	55	84.44	0.548	1.24	0.939	54.07% Precentral R
RH SomMot 4	16	2	0.13	0.544	4	1	138	18	17.25	0.591	1.50	0.530	62.76% Postcentral R
LH_SalVentAttn_ParOper_1	12	2	0.17	0.376	26	3	1157	98	99.17	0.601	1.50	0.371	54.68% SupraMarginal_L

LH SomMot 5	19	4	0.21	0.133	13	2	417	44	46.33	0.614	1.26	0.636	55.07% Postcentral_L
RH DorsAttn Post 5	25	2	0.08	0.800	6	4	246	166	51.79	0.632	1.20	0.800	46.68% Precuneus_R
LH SomMot 4	18	2	0.11	0.594	17	1	826	32	43.47	0.669	1.44	0.588	73.10% Postcentral_L
LH Default Temp 2	4	1	0.25	0.366	10	5	307	110	78.94	0.725	1.00	0.333	88.43% Temporal_Mid_L
RH DorsAttn Post 4	36	2	0.06	0.909	21	2	997	152	83.08	0.733	1.00	0.909	56.13% Parietal_Sup_R
RH DorsAttn Post 3	19	1	0.05	0.891	11	1	327	17	27.25	0.740	0.89	0.714	42.84% Parietal_Inf_R
RH Limbic OFC 1	4	1	0.25	0.376	10	1	355	44	27.31	0.744	1.00	0.346	28.24% Rectus_R
RH SomMot 1	21	1	0.05	0.913	18	1	763	17	36.33	0.789	0.86	0.785	61.86% Temporal_Sup_R
RH DorsAttn Post 2	17	1	0.06	0.861	15	2	606	49	48.48	0.794	0.71	0.856	85.28% Postcentral_R
LH SalVentAttn FrOperIns 2	2	1	0.50	0.219	12	1	534	44	31.41	0.801	1.00	0.216	64.87% Insula_L
RH Cont Par 1	40	1	0.03	0.991	38	9	1722	376	190.09	0.804	0.63	0.984	59.43% SupraMarginal_R
LH DorsAttn Post 2	13	1	0.08	0.774	18	2	792	59	44.00	0.908	0.77	0.760	29.06% Parietal_Inf_L
LH Default Par 1	12	1	0.08	0.758	12	1	391	18	18.62	0.928	0.92	0.627	69.08% Temporal_Mid_L
LH Vis 7	6	1	0.17	0.497	19	2	836	50	54.52	0.936	1.00	0.391	42.60% Temporal_Mid_L
RH SalVentAttn TempOccPar 2	39	1	0.03	0.988	25	1	1113	34	27.83	0.977	0.59	0.988	68.61% SupraMarginal_R
LH SomMot 1	21	1	0.05	0.928	23	1	905	26	23.21	0.987	0.57	0.928	77.45% Temporal_Sup_L
LH_DorsAttn_Post_3	8	1	0.13	0.591	38	5	1778	242	79.02	0.997	0.50	0.591	48.76% Parietal_Sup_L

**Table S6: Region-wise fNIRS meta-analyses**

Results from region-wise meta-analysis of fNIRS INS experiments, for the complete and for a restricted set of experiments. *Parcel*: brain region label drawn from a functionally defined 100-parcel cortical parcellation (21). *N (ch/exp/sub)*: number of channels, experiments, and subjects associated to the respective region. *Ratio*: (number of INS channels / number of all channels). *Ratio \* N (sub-all)*: (Ratio \* number of subjects contributing to a parcel). *Ratio \* N (exp-all)*: (Ratio \* number of experiments contributing to a parcel). *p (M)*: Median *p* value resulting from iterative (1,000 iterations) recalculation of fNIRS meta-analysis after fNIRS coordinate randomization within a 1 cm cortical radius. *p (%)*: Percentage of sub-threshold (< 0.05) *p* values after coordinate randomization. Randomization analysis was only performed for the main fNIRS coordinate dataset. *P* values are estimated from null distributions generated by permuting (5,000 iterations) the fNIRS channel coordinate-atlas parcel assignment and recalculating the respective index. *AAL region*: Anatomical region with the largest coverage from the automated anatomic labelling atlas (117). The order is based on *p* values associated to “*Ratio \* N (sub-all)*” and only those parcels in which at least one INS channel was observed are shown.

Abbreviations: fNIRS = functional near-infrared spectroscopy, INS = interpersonal neural synchronization.

Topic	rTPJ			Whole-brain					
	Reverse inference			Forward inference			Spearman correlation		
	q	z	prob.	q	z	lik.	p	q	Z(rho)
145 mind mental social	<0.001	6.81	0.02	<0.001	5.40	1.91	0.379	0.679	0.06
143 action actions observation	<0.001	5.32	0.02	<0.001	3.71	1.56	<0.001	0.005	0.64
82 motion mt moving	<0.001	5.25	0.01	<0.001	4.29	1.84	0.001	0.007	0.54
193 mirror video imitation	0.007	2.68	0.01	0.211	1.25	1.26	0.001	0.009	0.51
115 face faces fusiform	0.011	2.55	0.02	0.048	1.98	1.34	0.224	0.498	0.18
154 social interactions interaction	0.012	2.50	0.02	0.012	2.51	1.43	0.777	1.000	-0.19
138 real virtual reality	0.021	2.31	0.01	0.027	-2.21	1.54	0.012	0.075	0.44
30 events future personal	0.072	1.80	0.01	0.090	-1.70	1.45	0.838	1.000	-0.23
99 detection novelty oddball	0.072	1.80	0.01	0.025	-2.24	1.71	0.187	0.458	0.17
175 dopamine dopaminergic striatum	0.145	-1.46	0.00	0.997	0.00	0.58	1.000	1.000	-0.66
149 olfactory taste odor	0.204	-1.27	0.00	0.997	0.00	0.34	0.992	1.000	-0.46
100 gestures abstract race	0.243	1.17	0.01	0.210	-1.25	1.43	0.042	0.146	0.38
142 scene scenes perspective	0.243	1.17	0.01	0.217	-1.23	1.30	0.242	0.512	0.17
26 time delay temporal	0.243	1.17	0.02	0.217	1.23	1.17	0.381	0.679	0.07
64 attention attentional visual	0.244	1.16	0.03	0.217	1.23	1.14	<0.001	0.005	0.82
139 faces emotional facial	0.246	1.16	0.02	0.142	1.47	1.28	0.881	1.000	-0.22
157 somatosensory stimulation cortex	0.248	-1.15	0.01	0.997	0.00	0.65	0.394	0.689	0.07
191 eye gaze saccade	0.248	1.15	0.01	0.437	-0.78	1.17	0.059	0.180	0.35
97 adaptation selective stimulus	0.248	1.15	0.03	0.210	1.25	1.16	<0.001	0.005	0.64
8 cues cue target	0.252	1.14	0.01	0.335	0.96	1.17	0.041	0.146	0.35
181 creative creativity generation	0.261	1.12	0.00	0.185	-1.32	1.56	0.435	0.719	0.04
130 risk high taking	0.292	-1.05	0.00	0.997	0.00	0.72	0.998	1.000	-0.50
74 feedback negative performance	0.292	1.05	0.01	0.368	-0.90	1.22	0.470	0.750	0.03
11 cognitive function performance	0.292	-1.05	0.05	0.906	0.12	0.98	0.523	0.821	-0.02
108 visual auditory sensory	0.387	0.87	0.02	0.449	0.76	1.10	<0.001	0.005	0.61
111 memory encoding hippocampal	0.387	-0.87	0.01	0.997	0.00	0.84	0.835	1.000	-0.21
118 goal goals planning	0.387	-0.87	0.00	0.997	0.00	0.65	0.744	1.000	-0.15
125 expertise experts ic	0.387	-0.87	0.00	0.997	0.00	0.67	0.026	0.125	0.36
152 perceptual perception visual	0.387	-0.87	0.01	0.997	0.00	0.87	0.001	0.008	0.56
159 inhibition response inhibitory	0.387	0.87	0.01	0.217	1.23	1.23	0.461	0.749	0.03
161 women men sex	0.387	-0.87	0.01	0.997	0.00	0.81	0.999	1.000	-0.49
166 pictures picture images	0.387	-0.87	0.01	0.997	0.00	0.83	0.928	1.000	-0.20
184 spatial location space	0.387	0.87	0.02	0.476	0.71	1.09	0.010	0.073	0.48
198 repetition priming suppression	0.387	-0.87	0.01	0.997	0.00	0.84	0.057	0.180	0.36
3 task switching set	0.387	0.87	0.01	0.515	-0.65	1.11	0.100	0.281	0.27
39 light shed alertness	0.387	0.87	0.01	0.335	-0.96	1.27	0.688	1.000	-0.13
55 music musical pitch	0.387	-0.87	0.00	0.997	0.00	0.62	0.425	0.717	0.05
6 implicit explicit cd	0.387	0.87	0.01	0.216	-1.24	1.43	0.364	0.676	0.06
196 personality trait scores	0.433	-0.78	0.01	0.872	-0.16	0.96	0.998	1.000	-0.48
79 training practice trained	0.433	-0.78	0.01	0.997	0.00	0.85	0.338	0.641	0.11
187 speech auditory temporal	0.437	0.78	0.02	0.599	0.53	1.06	0.212	0.494	0.18
31 hearing deaf sign	0.442	0.77	0.00	0.416	-0.81	1.33	0.025	0.125	0.40
28 memory retrieval episodic	0.448	0.76	0.01	0.745	0.33	1.03	0.836	1.000	-0.23
197 reward striatum anticipation	0.466	-0.73	0.01	0.906	0.12	0.96	1.000	1.000	-0.58
117 object objects visual	0.484	0.70	0.01	0.449	0.76	1.12	0.030	0.125	0.42
58 autonomic arousal rate	0.484	-0.70	0.00	0.997	0.00	0.76	0.988	1.000	-0.44
20 wm load memory	0.491	-0.69	0.01	0.997	0.00	0.79	0.191	0.458	0.19
93 language sentences comprehension	0.491	0.69	0.01	0.449	0.76	1.12	0.102	0.281	0.28
128 prediction error outcome	0.500	0.67	0.01	0.281	1.08	1.21	0.762	1.000	-0.12
137 regulation emotion reappraisal	0.503	0.67	0.01	0.449	-0.76	1.16	0.997	1.000	-0.36
182 experience subjective ratings	0.503	0.67	0.01	0.416	0.81	1.14	0.967	1.000	-0.28
5 body bodies eba	0.503	0.67	0.01	0.335	-0.96	1.29	0.049	0.165	0.37
127 task performance cognitive	0.513	-0.65	0.14	0.821	0.23	1.00	0.029	0.125	0.34
158 pet glucose metabolism	0.520	0.64	0.00	0.772	-0.29	1.01	1.000	1.000	-0.56
95 verbs verb nouns	0.567	0.57	0.00	0.745	-0.33	1.03	0.056	0.180	0.34
186_decision_making_choice	0.625	0.49	0.02	0.449	0.76	1.11	0.811	1.000	-0.19

23 reasoning relational relations	0.628	0.48	0.00	0.452	-0.75	1.21	0.289	0.585	0.14
173 touch ct tactile	0.630	-0.48	0.00	0.997	0.00	0.72	0.010	0.073	0.47
43 conflict interference incongruent	0.630	0.48	0.01	0.519	0.64	1.10	0.023	0.125	0.40
146 pain painful chronic	0.657	-0.44	0.01	0.997	0.00	0.78	0.903	1.000	-0.29
18 color shape shapes	0.718	0.36	0.01	0.449	-0.76	1.20	0.001	0.007	0.55
57 learning learned sequence	0.718	-0.36	0.01	0.997	0.00	0.85	0.791	1.000	-0.17
90 imagery mental rotation	0.718	-0.36	0.00	0.997	0.00	0.74	0.093	0.272	0.33
150 reading phonological readers	0.730	-0.35	0.01	0.997	0.00	0.82	0.001	0.007	0.57
153 phase women menstrual	0.853	-0.19	0.00	0.916	-0.11	0.87	0.947	1.000	-0.33
172 verbal fluency overt	0.853	-0.19	0.01	0.997	0.00	0.81	0.030	0.125	0.37
179 memory working task	0.853	-0.19	0.01	0.957	0.05	0.94	0.137	0.346	0.24
2 problem problems arithmetic	0.853	-0.19	0.00	0.957	-0.05	0.88	0.134	0.346	0.26
52 control cognitive task	0.853	-0.19	0.01	0.821	0.23	0.99	0.323	0.625	0.11
121 tool tools knowledge	0.857	0.18	0.00	0.636	-0.47	1.11	0.035	0.136	0.41
13 fear conditioning extinction	0.857	0.18	0.00	0.732	-0.34	1.05	0.999	1.000	-0.44
131 test performance intelligence	0.857	0.18	0.01	0.599	-0.53	1.10	0.855	1.000	-0.23
160 bias biases spd	0.857	-0.18	0.00	0.821	-0.23	0.97	0.825	1.000	-0.14
163 language chinese english	0.857	-0.18	0.01	0.957	-0.05	0.90	0.010	0.073	0.47
167 human humans animal	0.857	0.18	0.01	0.817	0.23	1.00	0.814	1.000	-0.20
177 mood rumination induction	0.857	0.18	0.00	0.519	-0.64	1.18	0.983	1.000	-0.38
180 amygdala threat fear	0.857	-0.18	0.01	0.821	-0.23	0.99	0.998	1.000	-0.45
65 recognition correct familiarity	0.857	0.18	0.01	0.719	-0.36	1.05	0.304	0.602	0.11
9 semantic word knowledge	0.857	-0.18	0.01	0.965	0.04	0.92	0.286	0.585	0.13
42 negative positive vmcfc	0.891	0.14	0.01	0.599	0.53	1.07	1.000	1.000	-0.58
116 task matching strategy	0.912	-0.11	0.01	0.906	-0.12	0.95	0.015	0.092	0.42
91 movement motor movements	0.926	-0.09	0.01	0.997	0.00	0.74	0.221	0.498	0.22
92 executive control functions	0.926	-0.09	0.01	0.957	-0.05	0.91	0.403	0.693	0.06
183 orientation colour separation	0.959	-0.05	0.00	0.821	-0.23	0.97	0.238	0.512	0.17
162 words word lexical	0.960	-0.05	0.01	0.997	0.00	0.87	0.029	0.125	0.39
169 acupuncture stimulation vstm	0.960	-0.05	0.00	0.946	-0.07	0.82	0.816	1.000	-0.18
112 context empathy contextual	0.962	0.05	0.01	0.722	-0.36	1.05	0.759	1.000	-0.17
147 target search targets	0.963	-0.05	0.01	0.821	0.23	0.99	0.036	0.136	0.34
189 category categories categorization	0.974	0.03	0.01	0.745	-0.33	1.03	0.117	0.314	0.28
19 illusion physical perceived	0.974	0.03	0.00	0.772	-0.29	1.02	<0.001	0.007	0.53
66_emotional_negative_amygdala	0.988	0.02	0.02	0.775	0.29	1.01	0.998	1.000	-0.45

**Table S7: Functional-decoding results**

Functional decoding of the rTPJ cluster using reverse and forward inference estimates, and the whole-brain INS distributions using spatial Spearman correlations ( $Z$ -transformed), sorted by reverse inference  $q$  values. Topics are selected from 200 latent Dirichlet allocation topics distributed with the Neurosynth database (version 7). For whole-brain correlations,  $p$  values are derived from comparison to correlations with topic-wise null maps (10,000 permutations) and were false discovery rate-corrected across all topics ( $q$ ). All associations significant after multiple comparison correction in any analysis are marked in bold.

Abbreviations: rTPJ = right temporo-parietal junction, prob. = probability, lik. = likelihood.

**In-vivo & post-mortem neurotransmitter atlases**

Atlas	Whole-brain			Cortex-only		Baseline-adjusted	
	$Z(\rho)$	$p$	$q$	$Z(\rho)$	$p$	$Z(\rho)$	$p$
GABAa	0.51	<0.001	0.005	0.43	0.002	0.51	<0.001
SV2a	0.40	0.002	0.022	0.29	0.047	0.41	0.001
mGluR5	0.37	0.003	0.026	0.26	0.089	0.36	0.005
5HT2a	0.43	0.011	0.065	0.19	0.160	0.49	0.005
M1	0.27	0.032	0.154	0.28	0.063	0.26	0.040
5HT6	0.19	0.084	0.320				
5HT1a	0.27	0.093	0.320				
5HT1b	0.18	0.115	0.346				
NET	0.17	0.180	0.480				
NMDA	-0.06	0.590	1.000				
D1	-0.07	0.623	1.000				
D2	-0.12	0.697	1.000				
CB1	-0.08	0.726	1.000				
5HT4	-0.13	0.737	1.000				
a4b2	-0.16	0.755	1.000				
FDOPA	-0.19	0.782	1.000				
DAT	-0.19	0.788	1.000				
OXT	-0.19	0.791	1.000				
H3	-0.20	0.908	1.000				
MU	-0.23	0.929	1.000				
5HTT	-0.35	0.942	1.000				
VAcHT	-0.29	0.956	1.000				
OXTR	-0.39	0.984	1.000				
CD38	-0.57	1.000	1.000				

**GABA-related gene co-expression clusters**

Atlas	Whole-brain		
	$Z(\rho)$	$p$	$q$
Cluster 3	0.45	0.005	0.014
Cluster 2	0.43	0.007	0.014
Cluster 4	-0.19	0.932	0.952
Cluster 1	-0.23	0.952	0.952

**Table S8: Results of neurotransmitter association analyses**

Spatial correlations between interpersonal neural synchronization  $Z$ -maps and neurotransmitter receptor distributions derived from invivo nuclear imaging atlases (144, 145) or post-mortem mRNA expression data (19, 20). Correlation coefficients are r-to- $Z$  transformed partial Spearman's  $\rho$ ,  $p$  values are uncorrected and derived from null-correlations,  $q$  values are false discovery-corrected  $p$  values according to the Benjamini-Hochberg procedure. The two right main columns show sensitivity analyses of the significant positive associations (i) calculated only on cortical parcels and (ii) calculated after additional inclusion of functional baseline activation rate (BL) in partial correlations of INS and nuclear imaging maps. Rows are sorted by main  $p$  values.



**Neuronal cell types**

Category	N (genes)	Average Z (p)	p (norm)	q (norm)	p (perm)	q (perm)
Adult-Ex3	9	0.36	< 0.001	0.001	< 0.001	< 0.001
Adult-In5	7	0.12	0.001	0.013	0.001	0.012
Adult-In6	5	0.19	0.005	0.039	0.004	0.032
Adult-Ex4	15	0.12	0.030	0.147	0.025	0.125
Adult-OtherNeuron	38	0.07	0.031	0.147	0.026	0.125
Adult-Ex5	21	0.11	0.051	0.206	0.049	0.197
Adult-Ex2	9	0.06	0.095	0.325	0.095	0.326
Adult-Ex1	17	-0.01	0.566	1.000	0.563	1.000
Adult-Ex6	34	-0.11	0.993	1.000	0.994	1.000
Adult-Ex7	6	-0.07	0.908	1.000	0.906	1.000
Adult-Ex8	54	-0.07	0.931	1.000	0.928	1.000
Adult-In1	2	-0.30	0.994	1.000	0.993	1.000
Adult-In2	7	-0.10	0.991	1.000	0.993	1.000
Adult-In3	17	-0.02	0.751	1.000	0.751	1.000
Adult-In4	11	-0.04	0.788	1.000	0.791	1.000
Adult-In7	10	-0.03	0.759	1.000	0.762	1.000
Adult-In8	3	0.02	0.368	1.000	0.369	1.000
Adult-Astro	37	-0.23	1.000	1.000	1.000	1.000
Adult-Endo	76	-0.16	0.992	1.000	0.992	1.000
Dev-quiescent	15	-0.04	0.785	1.000	0.780	1.000
Dev-replicating	25	-0.05	0.971	1.000	0.973	1.000
Adult-Micro	20	-0.25	0.998	1.000	0.999	1.000
Adult-OPC	39	-0.19	0.999	1.000	0.999	1.000
Adult-Oligo	35	-0.03	0.649	1.000	0.651	1.000

**Developmental regional enrichment**

Category	N (genes)	Average Z (p)	p (norm)	q (norm)	p (perm)	q (perm)
child CBC	165	0.16	< 0.001	0.001	< 0.001	< 0.001
adolescent CBC	155	0.15	< 0.001	0.001	< 0.001	< 0.001
adult AIC	31	0.32	< 0.001	0.001	< 0.001	< 0.001
adult VIC	67	0.32	< 0.001	0.001	< 0.001	< 0.001
adult CBC	193	0.14	< 0.001	0.001	< 0.001	< 0.001
adolescent STC	5	0.50	< 0.001	< 0.001	< 0.001	0.001
adult STC	67	0.28	< 0.001	0.001	< 0.001	0.001
adult SIC	45	0.31	< 0.001	0.001	< 0.001	0.001
adult IPC	55	0.27	< 0.001	0.001	< 0.001	0.001
infant CBC	121	0.10	< 0.001	0.001	< 0.001	0.001
adult MIC	30	0.27	< 0.001	0.002	< 0.001	0.001
adolescent DFC	27	0.19	< 0.001	0.001	< 0.001	0.002
adolescent VFC	8	0.24	< 0.001	0.001	< 0.001	0.002
adult VFC	43	0.26	< 0.001	0.001	< 0.001	0.002
infant VIC	47	0.21	< 0.001	0.001	0.001	0.003
adolescent OFC	14	0.26	< 0.001	0.001	0.001	0.003
adolescent IPC	17	0.31	< 0.001	0.001	0.001	0.004
adult OFC	41	0.25	< 0.001	0.001	0.001	0.004
infant ITC	8	0.19	0.001	0.005	0.001	0.005
adult DFC	57	0.21	0.001	0.005	0.002	0.007
adolescent VIC	11	0.18	0.002	0.008	0.002	0.007
infant IPC	16	0.21	0.002	0.008	0.003	0.010
infant MFC	8	0.18	0.004	0.013	0.004	0.014
adult ITC	37	0.17	0.004	0.014	0.006	0.019
child VIC	31	0.12	0.006	0.019	0.006	0.019
infant SIC	35	0.15	0.008	0.024	0.007	0.022
prenatal VIC	171	0.07	0.009	0.025	0.008	0.024
adolescent SIC	9	0.15	0.010	0.028	0.009	0.025
adolescent ITC	15	0.13	0.010	0.028	0.010	0.026

**DisGeNET psychiatric disease markers**

Category	N (genes)	Average Z (p)	p (norm)	q (norm)	p (perm)	q (perm)
Seasonal Affective Disorder	14	0.200	< 0.001	0.001	< 0.001	< 0.001

Psychosis, Brief Reactive	11	0.238	< 0.001	0.001	< 0.001	< 0.001
Schizophreniform Disorders	11	0.238	< 0.001	0.001	< 0.001	< 0.001
Language Development Disorders	11	0.305	< 0.001	0.001	< 0.001	0.006
Speech Delay	11	0.305	< 0.001	0.001	< 0.001	0.006
Semantic-Pragmatic Disorder	11	0.305	< 0.001	0.001	< 0.001	0.006
Neurodevelopmental Disorders	89	0.151	< 0.001	0.001	< 0.001	0.006
Schizoaffective Disorder	25	0.136	< 0.001	0.003	< 0.001	0.006
Abnormal behavior	15	0.173	< 0.001	0.003	< 0.001	0.006
Global developmental delay	125	0.081	< 0.001	0.006	< 0.001	0.006
Major Affective Disorder 2	13	0.156	< 0.001	0.003	0.001	0.011
Opioid-Related Disorders	5	0.128	0.001	0.012	0.003	0.022
Opioid abuse	5	0.128	0.001	0.012	0.003	0.022
Narcotic Abuse	5	0.128	0.001	0.012	0.003	0.022
Opiate Addiction	5	0.128	0.001	0.012	0.003	0.022
Narcotic Dependence	5	0.128	0.001	0.012	0.003	0.022
Opiate Abuse	5	0.128	0.001	0.012	0.003	0.022
Manic Disorder	65	0.061	0.003	0.025	0.003	0.022
Phencyclidine Abuse	7	0.146	0.004	0.025	0.003	0.023
Phencyclidine-Related Disorders	7	0.146	0.004	0.025	0.003	0.023
Depression, Postpartum	6	0.189	0.004	0.025	0.004	0.027
Intellectual Disability	416	0.047	0.004	0.026	0.004	0.027
Developmental Coordination Disorder	7	0.117	0.005	0.034	0.005	0.031
Motor Skills Disorders	7	0.117	0.005	0.034	0.005	0.031
Profound Mental Retardation	126	0.041	0.008	0.043	0.006	0.036
Mental Retardation, Psychosocial	126	0.041	0.008	0.043	0.006	0.036
Mental deficiency	126	0.041	0.008	0.043	0.006	0.036
Manic	71	0.048	0.010	0.054	0.008	0.045

### Gene ontology biological processes

Category	N (genes)	Average Z ( $\rho$ )	p (norm)	q (norm)	p (perm)	q (perm)
neg. reg. of voltage-gated calcium channel activity	5	0.23	< 0.001	0.008	< 0.001	< 0.001
pos. reg. of skeletal muscle cell differentiation	5	0.42	< 0.001	0.008	< 0.001	< 0.001
reg. of water loss via skin	9	0.21	< 0.001	0.011	< 0.001	< 0.001
pos. reg. of nonmotile primary cilium assembly	6	0.24	< 0.001	0.011	< 0.001	< 0.001
deadenylation-dependent decapping of nuclear-transcribed mRNA	8	0.28	< 0.001	0.011	< 0.001	< 0.001
protein K6-linked ubiquitination	7	0.26	< 0.001	0.011	< 0.001	< 0.001
reg. of macromitophagy	5	0.28	< 0.001	0.011	< 0.001	< 0.001
cerebellar Purkinje cell differentiation	10	0.23	< 0.001	0.011	< 0.001	< 0.001
reg. of Golgi to plasma membrane protein transport	7	0.37	< 0.001	0.011	< 0.001	< 0.001
reg. of mRNA splicing, via spliceosome	65	0.14	< 0.001	0.011	< 0.001	< 0.001
phosphorelay signal transduction system	7	0.26	< 0.001	0.011	< 0.001	< 0.001
cardiac muscle adaptation	17	0.16	< 0.001	0.011	< 0.001	< 0.001
muscle hypertrophy in response to stress	16	0.19	< 0.001	0.011	< 0.001	< 0.001
cardiac muscle hypertrophy in response to stress	16	0.19	< 0.001	0.011	< 0.001	< 0.001
response to parathyroid hormone	7	0.24	< 0.001	0.011	< 0.001	< 0.001
reg. of protein sumoylation	21	0.14	< 0.001	0.011	< 0.001	< 0.001
cell differentiation in hindbrain	16	0.17	< 0.001	0.013	< 0.001	< 0.001
reg. of Rac protein signal transduction	12	0.21	< 0.001	0.013	0.001	0.020
histone H2A monoubiquitination	12	0.21	< 0.001	0.013	< 0.001	< 0.001
N-terminal protein lipidation	6	0.30	< 0.001	0.013	< 0.001	< 0.001
nephric duct development	8	0.19	< 0.001	0.013	< 0.001	0.010
neg. reg. of glucocorticoid receptor signaling pathway	6	0.34	< 0.001	0.013	< 0.001	0.010
histone H4-K5 acetylation	16	0.17	< 0.001	0.013	< 0.001	< 0.001
histone H4-K8 acetylation	16	0.17	< 0.001	0.013	< 0.001	< 0.001
cerebellar Purkinje cell layer formation	11	0.19	< 0.001	0.013	< 0.001	< 0.001
reg. of skeletal muscle cell differentiation	14	0.15	< 0.001	0.013	< 0.001	< 0.001
neg. reg. of mitophagy	6	0.31	< 0.001	0.013	< 0.001	< 0.001
reg. of alternative mRNA splicing, via spliceosome	35	0.13	< 0.001	0.013	< 0.001	0.015
reg. of production of miRNAs involved in gene silencing by miRNA	9	0.15	< 0.001	0.013	< 0.001	< 0.001
serine phosphorylation of STAT3 protein	6	0.35	< 0.001	0.013	< 0.001	< 0.001

pyrimidine nucleotide catabolic process	11	0.15	< 0.001	0.013	< 0.001	0.010
Golgi ribbon formation	11	0.15	< 0.001	0.013	< 0.001	0.010
startle response	24	0.14	< 0.001	0.013	< 0.001	< 0.001
histone deubiquitination	22	0.16	< 0.001	0.013	< 0.001	0.010
neg. reg. of peptidyl-tyrosine phosphorylation	32	0.13	< 0.001	0.013	< 0.001	< 0.001
establishment of meiotic spindle localization	5	0.19	< 0.001	0.013	< 0.001	< 0.001
pos. reg. of circadian rhythm	13	0.14	< 0.001	0.013	< 0.001	< 0.001
serine phosphorylation of STAT protein	8	0.23	< 0.001	0.013	< 0.001	< 0.001
locomotion involved in locomotory behavior	7	0.15	< 0.001	0.013	< 0.001	< 0.001
histone H4-K12 acetylation	7	0.22	< 0.001	0.013	< 0.001	0.010
macromitophagy	17	0.26	< 0.001	0.013	< 0.001	0.010
neg. reg. of histone H3-K9 methylation	10	0.21	< 0.001	0.013	< 0.001	< 0.001
cerebellar Purkinje cell layer morphogenesis	14	0.18	< 0.001	0.013	< 0.001	< 0.001
establishment of skin barrier	7	0.18	< 0.001	0.014	< 0.001	0.010
mRNA 3'-splice site recognition	6	0.26	< 0.001	0.014	< 0.001	0.015
reg. of circadian rhythm	82	0.10	< 0.001	0.014	< 0.001	0.015
response to muscle inactivity involved in reg. of muscle adaptation	5	0.23	< 0.001	0.014	< 0.001	< 0.001
response to denervation involved in reg. of muscle adaptation	5	0.23	< 0.001	0.014	< 0.001	< 0.001
reg. of type B pancreatic cell development	7	0.39	< 0.001	0.014	0.001	0.020
protein desumoylation	8	0.20	< 0.001	0.014	< 0.001	< 0.001

**Table S9: Results of gene category enrichment analyses**

Gene category enrichment analyses were calculated using ABAnnotate (32) on different databases; neuronal cell type markers (146–148), region-wise developmental mRNA expression data (149), psychiatric disease markers (150) and Gene Ontology biological process annotations (151, 152). The method was developed by Fulcher et al. (153). Spatial Spearman correlations were calculated between the interpersonal neural synchrony (INS) Z-map and the gene expression profiles drawn from the Allen Brain Atlas (19) of each gene annotated to a category. Correlation coefficients were averaged for each category. This process was repeated for 5,000 null maps derived from the INS map and significance was evaluated by comparison of the “real” category-wise average correlation with the 5,000 null correlations ( $p(\text{perm})$ ). To approximate p-values below 1/5000, a Gaussian distribution of category-wise average correlations was fitted to the distribution of permutation-derived scores ( $p(\text{norm})$ ). Resulting  $p$  values were false discovery rate-corrected according to the Benjamini-Hochberg procedure ( $q$ ). Rows are sorted according to the parametric uncorrected  $p$  values. Shown are all results for cell types, only significant for regional developmental enrichment, and the top 50 Gene Ontology categories. Full results are provided in the GitHub repository accompanying this publication.

Abbreviations: perm = permutation, norm = Gaussian normal distribution, CBC = cerebellar cortex, A1C = primary auditory cortex, V1C = primary visual cortex, STC = posterior (caudal) superior temporal cortex, S1C = primary somatosensory cortex, IPC = posteroventral (inferior) parietal cortex, M1C = primary motor cortex, DFC = dorsolateral prefrontal cortex, VFC = ventrolateral prefrontal cortex, OFC = orbital frontal cortex, ITC = inferolateral temporal cortex, MFC = anterior (rostral) cingulate (medial prefrontal) cortex, reg = regulation, pos = positive, neg = negative.

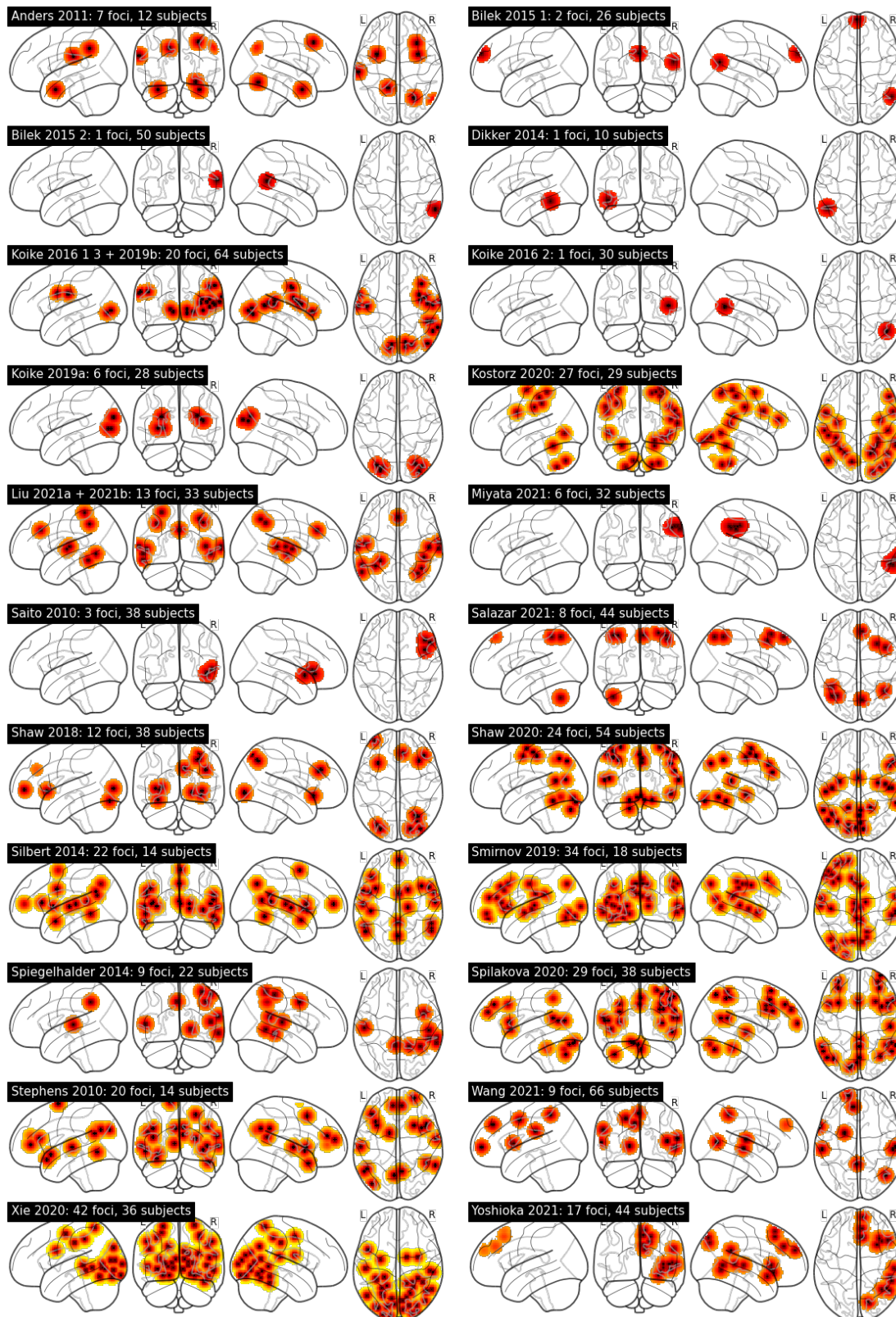
Cluster ID	N (terms)	Representative GO term		p (norm)	Average Z (p)
		GO term ID	GO term description		
1	16	GO:1901386	negative regulation of voltage-gated potassium channel activity	2.120E-06	0.23
2	49	GO:2001016	positive regulation of skeletal muscle cell differentiation	2.370E-06	0.42
3	36	GO:0033561	regulation of water loss via skin	8.690E-06	0.21
4	80	GO:0000290	deadenylation-dependent decapping of nuclear-transcribed mRNA	1.180E-05	0.28
5	13	GO:0085020	protein K6-linked ubiquitination	1.210E-05	0.26
6	18	GO:1901524	regulation of mitophagy	1.280E-05	0.28
7	26	GO:0021702	cerebellar Purkinje cell differentiation	1.330E-05	0.23
8	19	GO:0071107	response to parathyroid hormone	2.450E-05	0.24
9	30	GO:0033233	regulation of protein sumoylation	2.470E-05	0.14
10	21	GO:0006498	N-terminal protein lipidation	4.560E-05	0.30
11	9	GO:2000323	negative regulation of glucocorticoid receptor signaling pathway	4.700E-05	0.34
12	15	GO:0051295	establishment of meiotic spindle localization	7.030E-05	0.19
13	21	GO:0042501	serine phosphorylation of STAT protein	7.190E-05	0.23
14	5	GO:0031987	locomotion involved in locomotory behavior	7.250E-05	0.15
15	31	GO:0000389	mRNA 3'-splice site recognition	9.360E-05	0.26
16	13	GO:0014894	response to denervation involved in regulation of muscle adaptation	1.003E-04	0.23
17	16	GO:0030213	hyaluronan biosynthetic process	1.069E-04	0.19
18	6	GO:0032042	mitochondrial DNA metabolic process	1.802E-04	0.16
19	2	GO:0086027	AV node cell to bundle of His cell signaling	1.835E-04	0.30
20	6	GO:0000301	retrograde transport, vesicle recycling within Golgi	2.339E-04	0.13
21	19	GO:0015851	nucleobase transport	2.574E-04	0.34
22	2	GO:0042118	endothelial cell activation	3.987E-04	0.17
23	1	GO:0010659	cardiac muscle cell apoptotic process	6.881E-04	0.08
24	1	GO:0008228	opsonization	1.870E-03	0.21
25	2	GO:0039703	RNA replication	1.968E-03	0.08
26	1	GO:0039694	viral RNA genome replication	1.968E-03	0.08
27	2	GO:0086003	cardiac muscle cell contraction	3.916E-03	0.07

**Table S10: Results of GeneOntology category clustering**

The data were generated using GO-Figure! (154). GO categories significantly associated to the interpersonal neural synchrony Z-map (see Table S9) were clustered based on semantic similarity (similarity threshold  $\geq .2$ ) and for each cluster a representative term was selected in a data-driven fashion. *N (terms)* shows the number of GO categories annotated to each cluster. The list is ordered according to the *p* values of the representative terms and shows only GO terms that could be clustered. See Table S9 for further descriptions.

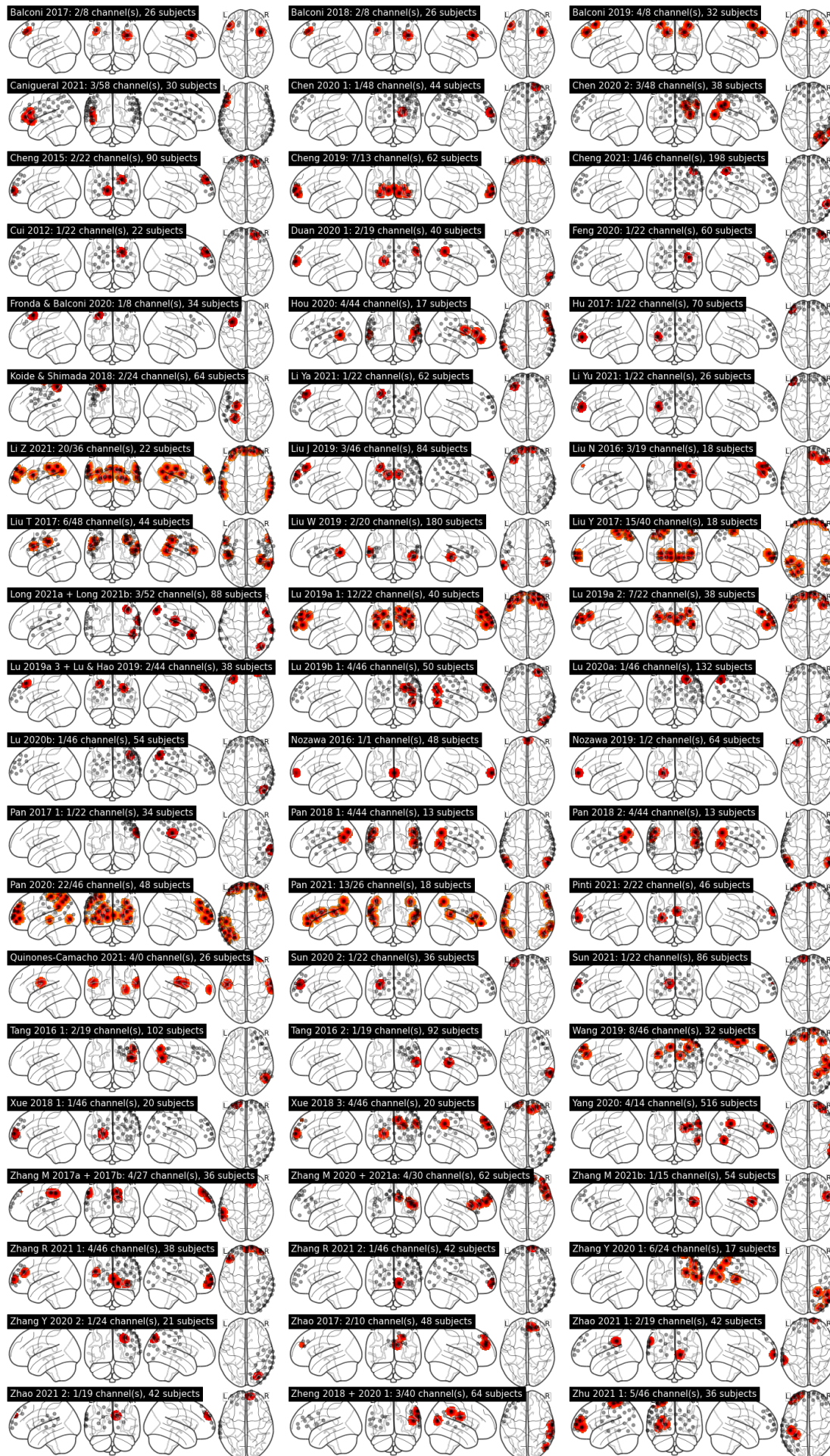
Abbreviations: GO = GeneOntology.

## 5 Supplementary Figures



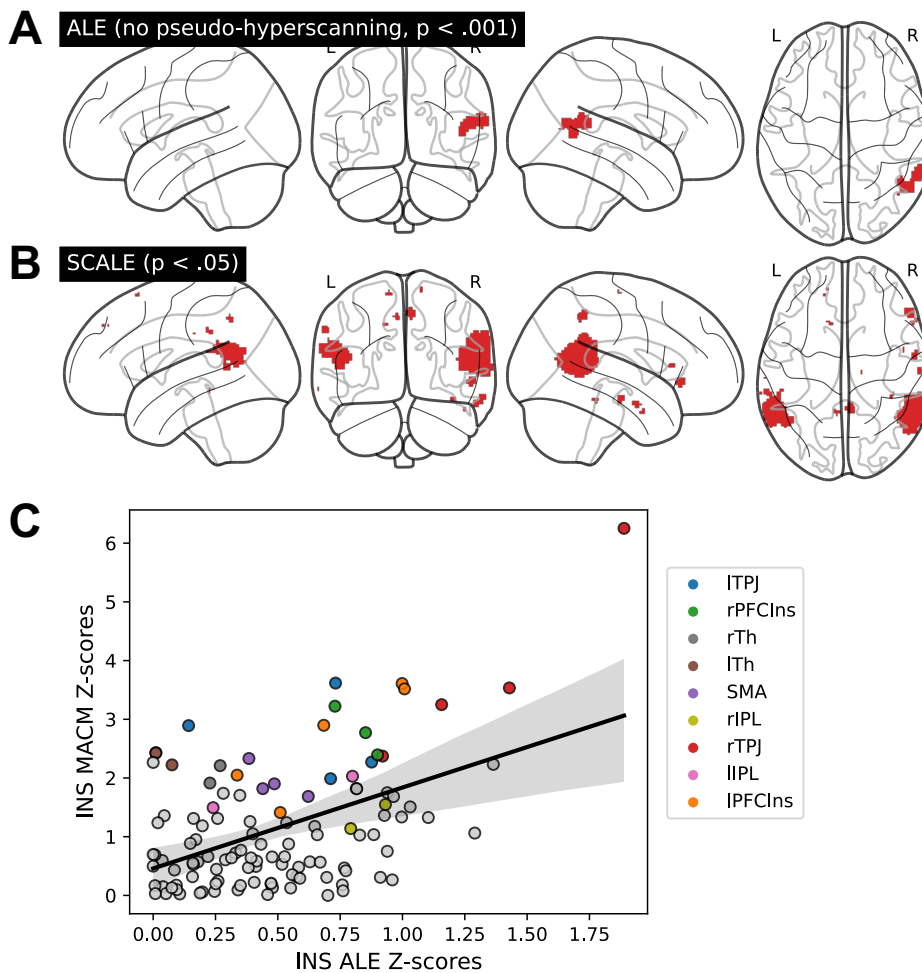
**Figure S1: INS foci reported in each fMRI experiment**

Foci are plotted individually for each experiment and smoothed using the activation likelihood estimation kernel depending on the reported sample size.



**Figure S2: INS foci reported in each fNIRS experiment**

Interpersonal neural synchrony (INS) channels are plotted individually for each experiment and, for visualization purposes, smoothed using the activation likelihood estimation kernel set to a fixed “sample size” of  $n = 10$ . Gray markers show all channels reported by, or reconstructed for, each study. Only those experiments reporting at least one INS channel are shown.

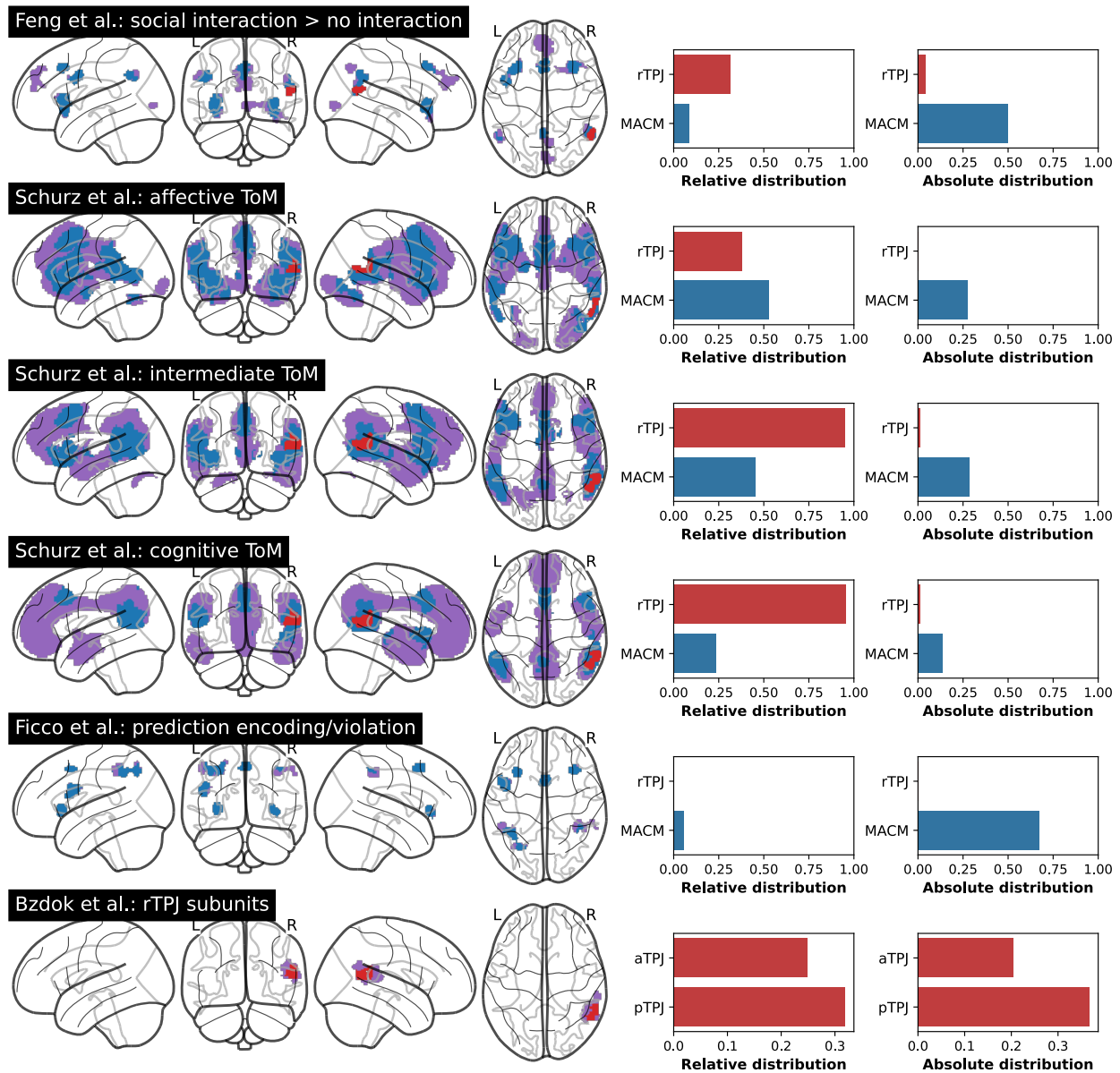


**Figure S3: ALE and MACM sensitivity analyses**

**A:** ALE without pseudo-hyperscanning studies. **B:** MACM corrected for baseline activation (SCALE). The method as implemented in NiMARE (158) does not support multiple comparison correction and was thresholded at voxel-level  $p < .05$ , uncorrected, for demonstration purposes. **C:** Spatial correlation between whole-brain distributions of INS and the rTPJ-associated MACM map to demonstrate comparable distributions patterns going beyond rTPJ activation.

Abbreviations: ALE = activation likelihood estimation, SCALE = specific coactivation likelihood estimation, INS = interpersonal neural synchronization, MACM = meta-analytic coactivation modeling, r/ITPJ = right/left temporoparietal junction, rSTG = right superior temporal gyrus, rIns = right insula, r/IPFCIns = right/left prefrontal cortex-insula, SMA = supplementary motor area, r/lTh = left thalamus, r/lIPL = left inferior parietal lobule, lPrec = left precuneus.

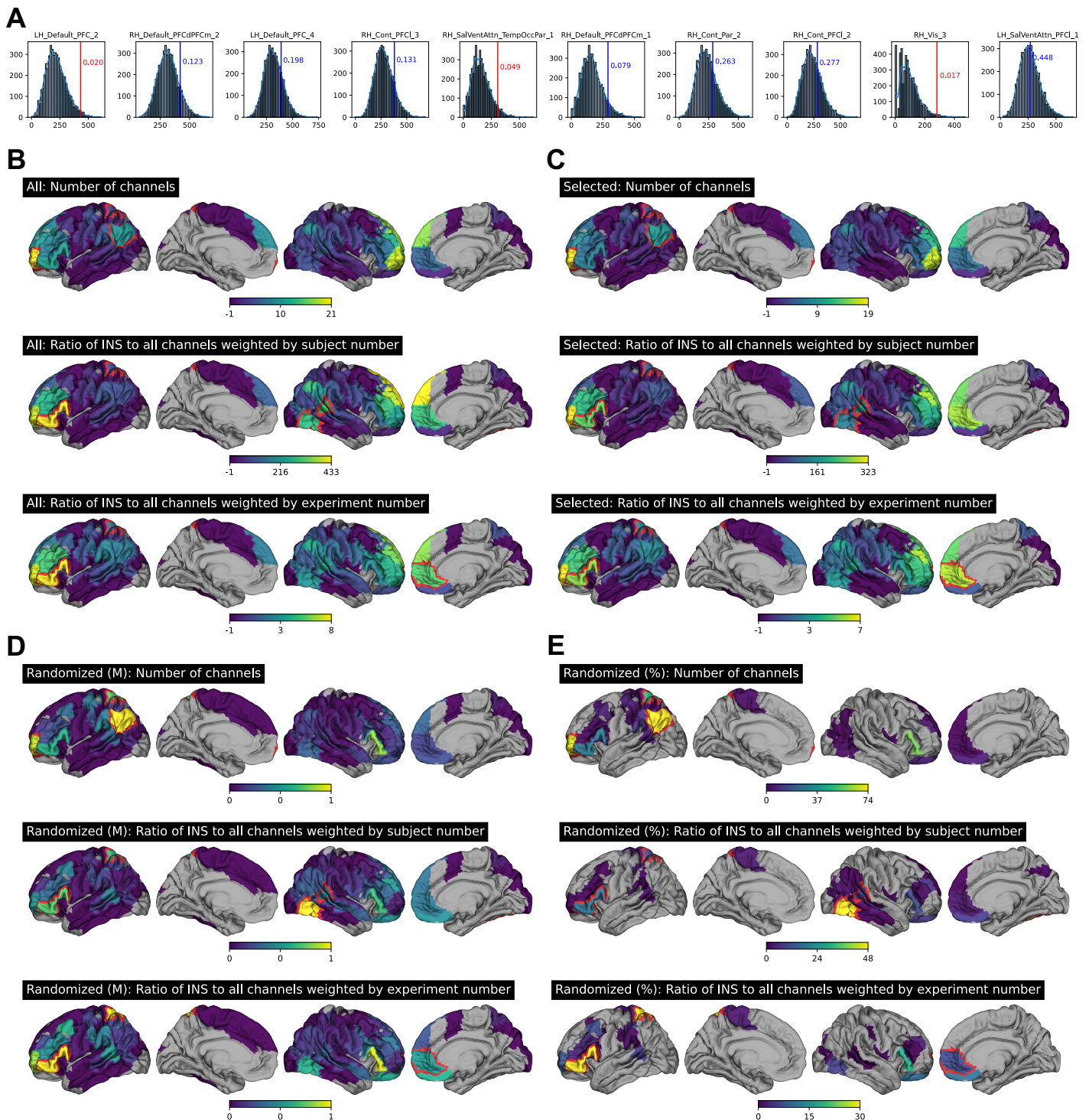




**Figure S4: Spatial overlap with prior meta-analytically derived networks and rTPJ subunits**

Sources: Feng et al., 2021 (159); Schurz et al., 2021 (160); Ficco et al., 2021 (161); Bzdok et al., 2013 (162). Original networks are plotted in purple, overlaid by conjunctions with the rTPJ-MACM network (blue) and the rTPJ cluster (red). For the upper five rows, the right side shows relative and absolute distributions of rTPJ cluster and MACM network within the corresponding meta-analytically derived network. The lowest row shows distributions of the INS-rTPJ cluster within two subunits of the rTPJ. Relative distribution: proportion of "INS-voxels" within a given network vs. all "INS-voxels". Absolute distribution: proportion of "INS-voxels" within a given network vs. all voxels within the network.

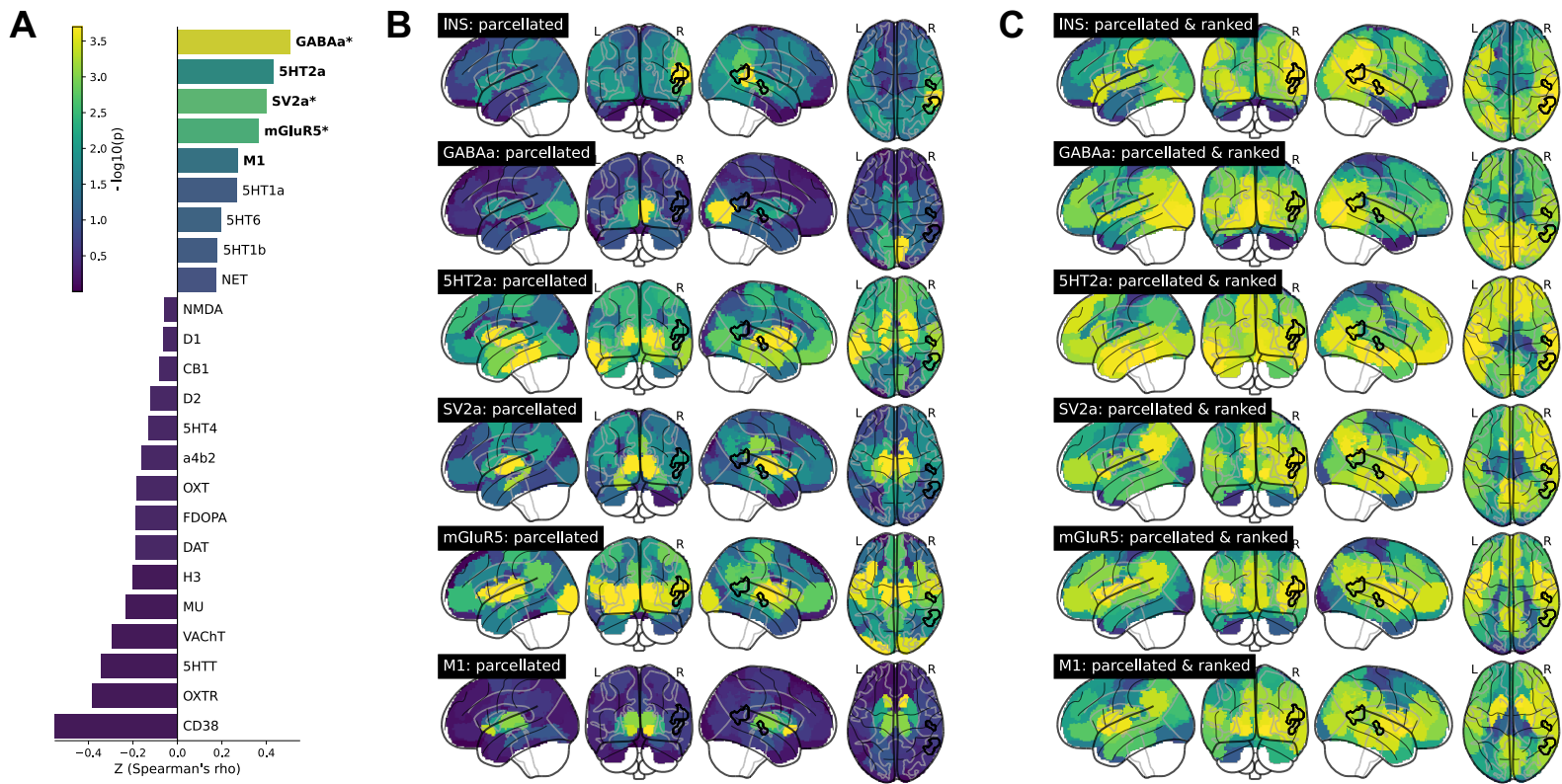
Abbreviations: rTPJ = right temporoparietal junction, MACM = meta-analytic connectivity modeling, ToM = theory of mind, INS = interpersonal neural synchronization.



**Figure S5: Complete meta-analytic results of INS fNIRS experiments**

**A:** Histograms for the top ten regions resulting from the permutations regarding the meta-analytic fNIRS INS analysis that included all experiments and focused on the parcel-wise ratio of INS vs. all channels weighted by the number of subjects. Vertical lines and numbers show the “real” result and the uncorrected  $p$  value estimated from each null distribution. **B:** Full fNIRS meta-analysis results for all included experiments. The middle row is shown in main Figure 2B. **C:** Full fNIRS meta-analysis results restricted to experiments explicitly assessing INS compared to rest, control, or randomization. **D:** Median (M)  $\log_{10}$ -transformed  $p$  values resulting from fNIRS meta-analysis after repeated randomization of fNIRS coordinates (radius = 1 cm, 1,000 iterations). **E:** Percentage of sub-threshold  $p$  values after coordinate randomization.

Abbreviations: INS = interpersonal neural synchrony, fNIRS = functional near-infrared spectroscopy.

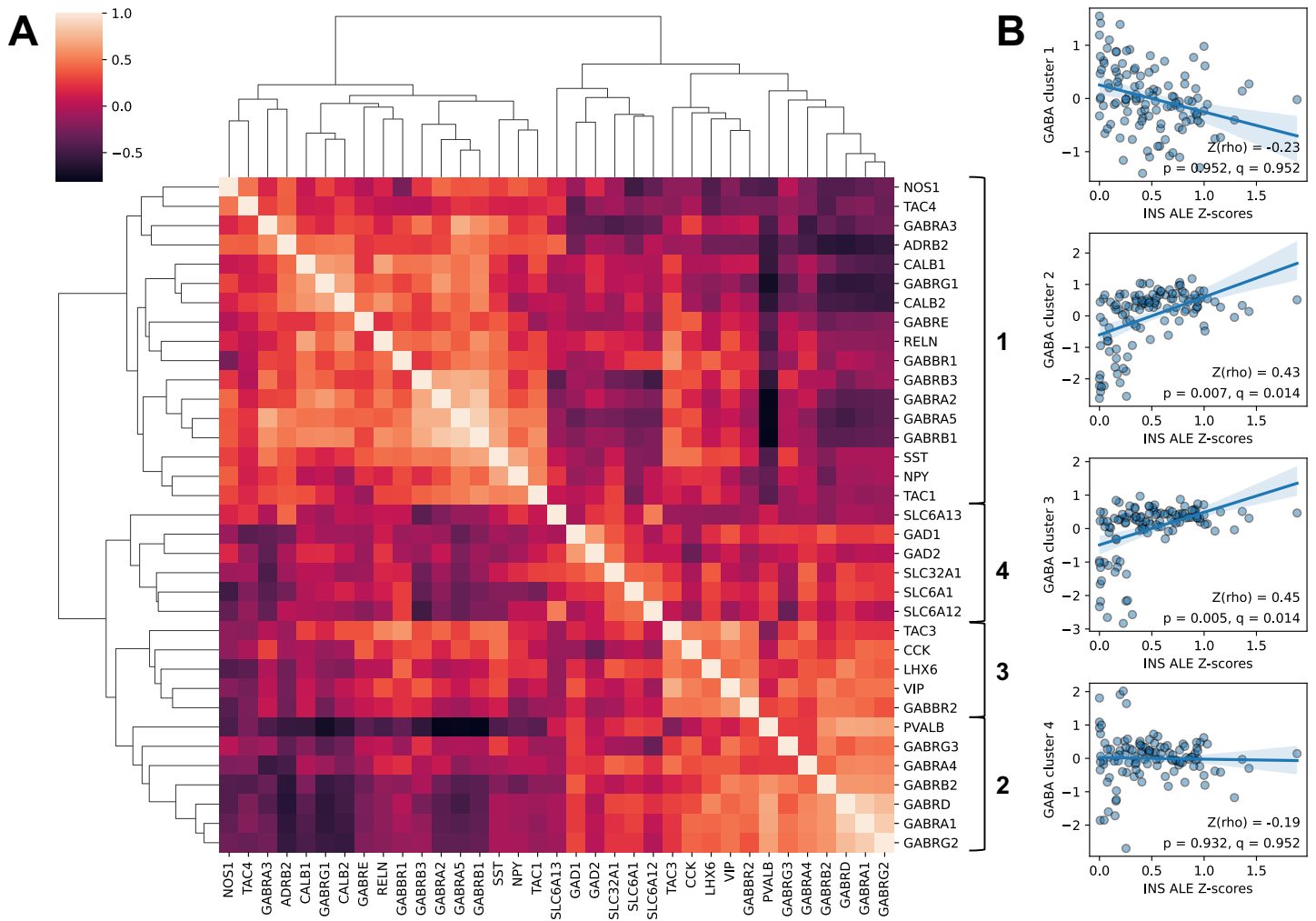


**Figure S6: Spatial associations of INS to neurotransmitter distributions**

**A:** Spatial correlations between whole-brain INS distribution and in vivo neurotransmitter receptor/ postmortem gene expression distributions. Color-coded  $p$  values are uncorrected, correlations significant at uncorrected  $p < .05$  derived from nonparametric permutation are printed in bold,  $p$  values surviving false discovery rate correction ( $q < .05$ ) are marked by asterisks. Nuclear imaging-derived maps were correlated with the INS map using partial Spearman correlations controlling for local gray matter volume to account for partial volume effects. Correlation coefficients were compared to those calculated from 5,000 spatial autocorrelation-preserving null maps derived from each individual transmitter map using JuSpyce. **B:** Parcellated and Z-transformed PET maps significantly associated to INS. **C:** To better demonstrate the spatial alignment between INS and PET maps, parcellated values were ranked and the ranks were reassigned to the brain volumes (yellow = highest rank). Black outline shows significant clusters resulting from the main activation likelihood estimation analysis.

See Table S3 for full neurotransmitter receptor names, nuclear imaging tracers, and template sources, and Table S8 for full JuSpyce results.

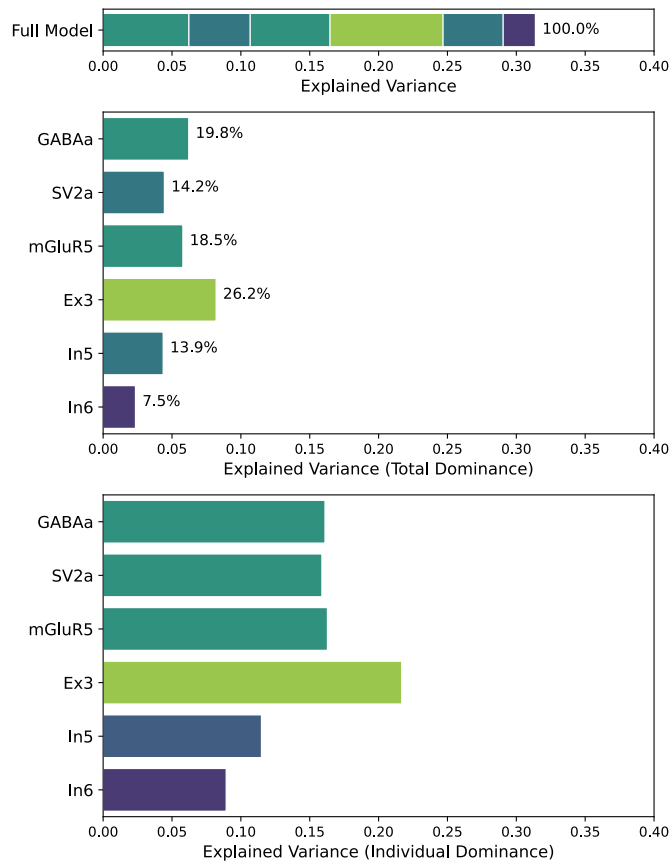
Abbreviations: INS = interpersonal neural synchrony, PET = positron emission tomography.



**Figure S7: Validation of INS-GABA<sub>A</sub> associations using mRNA expression data**

**A:** Clustered correlation heatmap of expression profiles of GABA-related genes. Colors represent Spearman correlations. Coexpression clusters were estimated using a hierarchical clustering algorithm based on Euclidean pairwise distances. **B:** Correlation between the INS distribution and gene cluster-wise averaged expression values; the correlations with clusters 2 and 3 are significant ( $q < .05$ ).

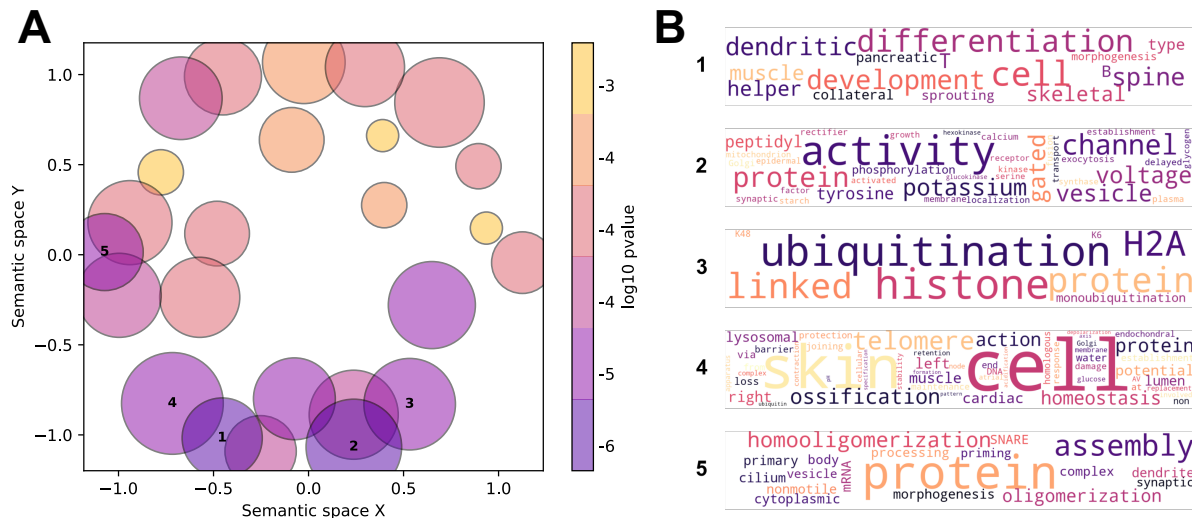
Abbreviations: INS = interpersonal neural synchronization, ALE = activation likelihood estimation.



**Figure S8: Variance of the INS distribution explained by significantly associated nuclear imaging and neuronal cell type data**

Results from a dominance analysis using significantly positively associated nuclear imaging and neuronal cell type atlases as predictors and the INS Z-map as target. As in the main univariate correlation analyses, local gray matter volume was regressed out of both predictors and target before performing the dominance analysis. Upper:  $R^2$  of the complete multilinear regression model ( $R^2 = .31$ ). Middle: “Total dominance” which can be interpreted as the relative contribution of a predictor to the explained variance of the complete model in interaction with the remaining predictors and therefore can also be expressed in percent of total  $R^2$ . Lower: “Individual dominance” which can be interpreted as each individual predictor’s explained variance independent of all other predictors.

Abbreviations: INS = interpersonal neural synchronization.



**Figure S9: Clustering of GeneOntology biological process categories spatially associated with INS**

**A:** Two-dimensional reduction of the pairwise semantic similarity matrix of 474 Gene Ontology (GO) categories spatially associated to INS. Each point represents one cluster of  $n$  biological process categories formed by a similarity threshold of .2 with the point size depicting  $n$ . Point color represents  $\log(p)$  associated with a representative GO term identified for each cluster. **B:** Word clouds built from combined category descriptions of the five semantic clusters most strongly associated to INS (marked by numbers 1-5 in both panels). Word size is based on word frequency within each semantic cluster.