





# Neuroimaging Spatial Colocalization Environment

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## Introduction

Most neuroimaging research focuses on **brain-regional effect estimates** as outcomes of interest.

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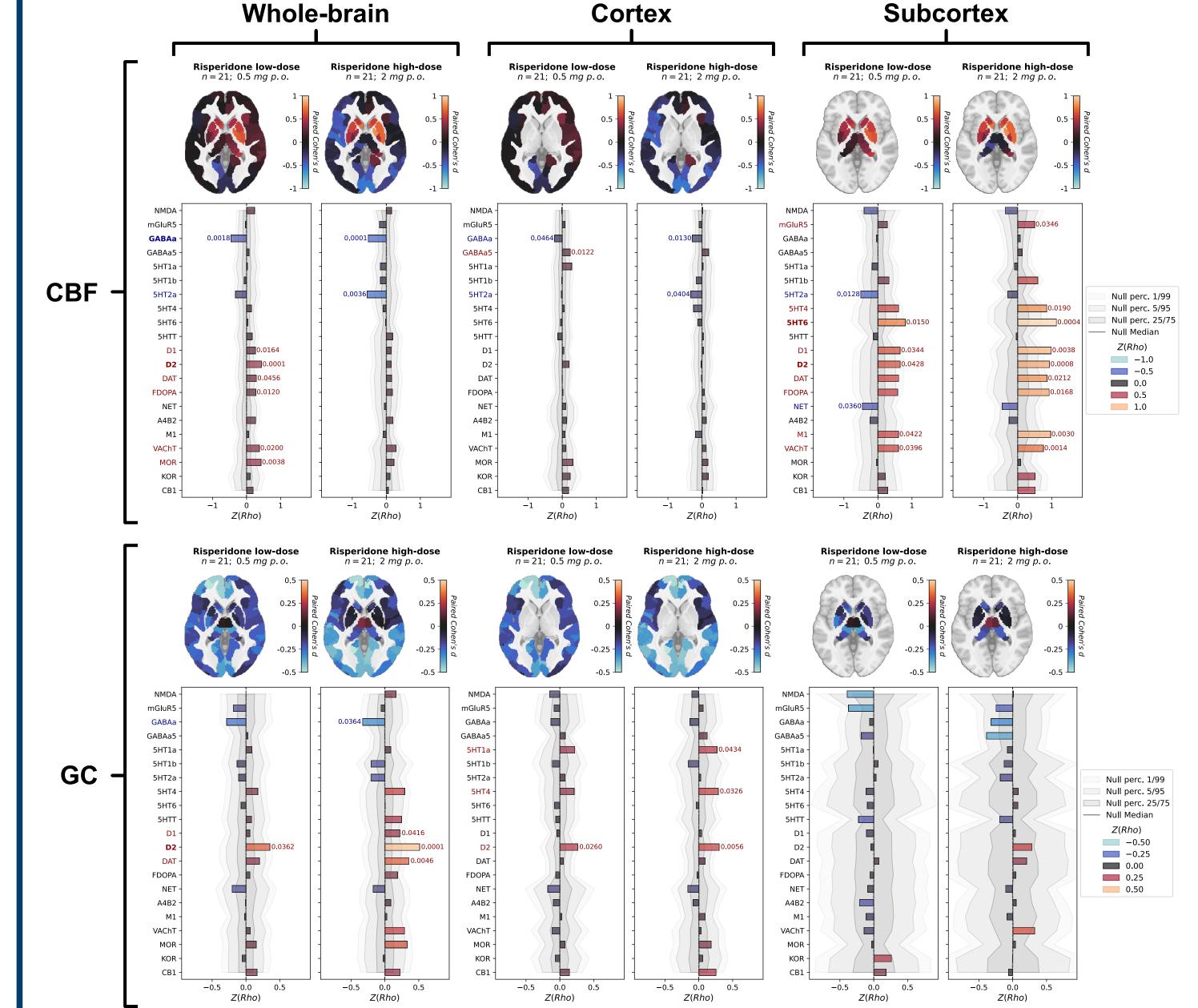
- Spatial patterns across the whole brain, cortex, or subcortex may convey important information missed by regionally constrained analysis approaches.
- These patterns are found in any given brain map, from **individual neurodevelopment** over time [1] to effect size maps reflecting **case-control** differences on the individual [2] or group level [3].
- By testing a **target brain map** for its **colocalization** with **reference atlases** of known biological or functional entities (e.g., neurotransmitters, gene expression, or meta-analytic maps of cognitive functions), one can derive **interpretable** inferences from in-vivo human neuroimaging data [1-5].

## Case studies

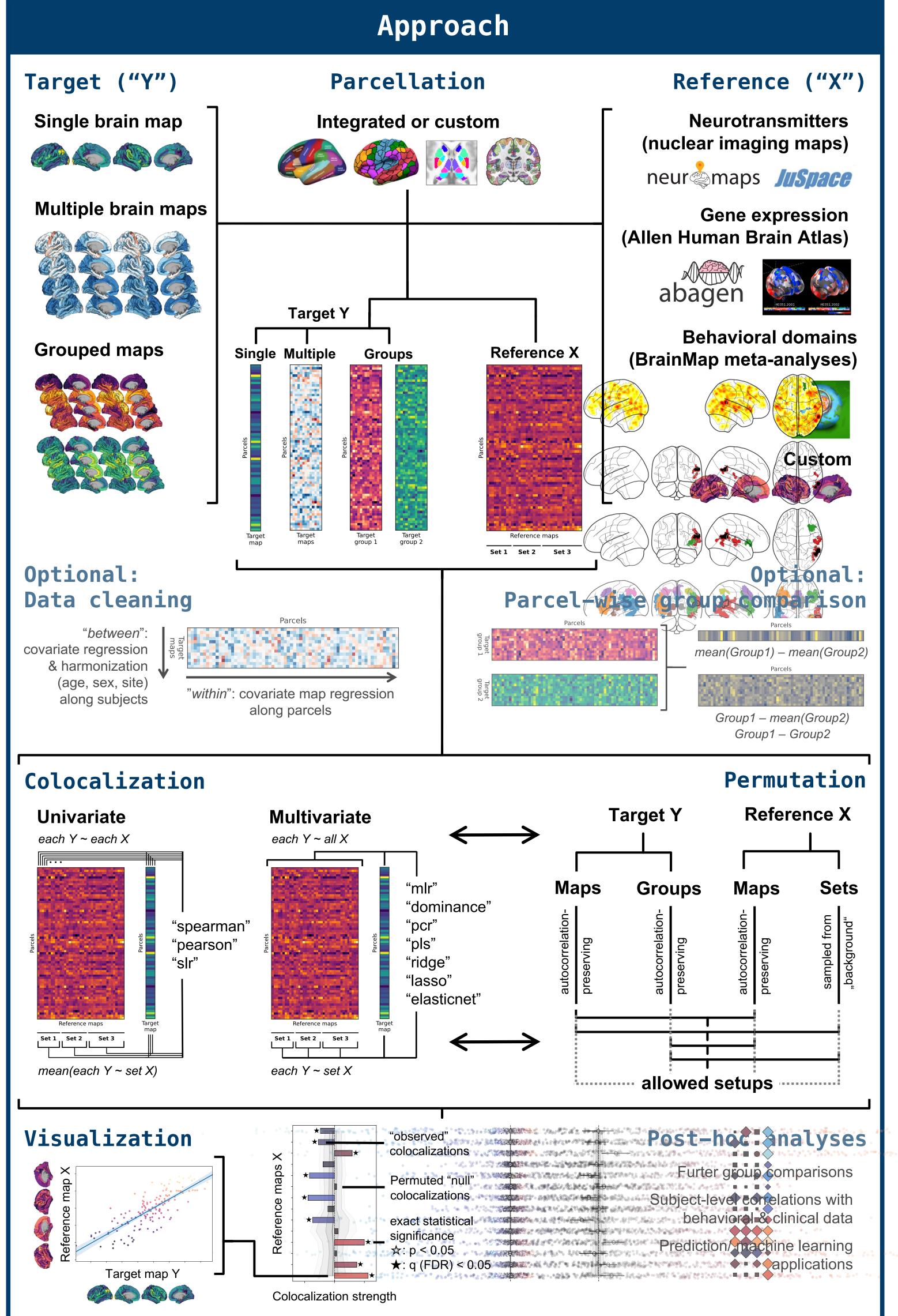
### **Drug challenge – Session permutation**

Colocalization between placebo-controlled drug effects on MRI (cerebral blood flow, CBF, and global correlation, GC) and independent neurotransmitter reference maps.

**Target**: CBF/GC maps [11] | **Reference**: group-average nuclear imaging maps (included in **NiSpace**) | **Parcellation**: Schaefer-200/ Melbourne-S2 | **Processing**: gray matter (GM) regression across parcels | **Comparison**: group level: parcel-wise paired Cohen's *d* | Colocalization: Spearman's Rho | Permutation: drug vs. placebo labels, shuffled within each subject, retaining constant 50/50 drug/placebo splits.



- **NiSpace** is a user-friendly API-focused toolbox integrated in the Python neuroimaging ecosystem that allows for fast, flexible, and easily scalable colocalization analysis.
- **NiSpace aims to provide high-level tools and workflows** for both novice and experienced users, starting with preprocessed volumetric and surface neuroimaging data, over data cleaning and harmonization, as well as several statistics and different permutation procedures, to results visualization.
- **NiSpace** applies and advances methods provided or introduced by other related toolboxes, e.g.: JuSpace [2], neuromaps [4], ENIGMA toolbox [5], BrainSpace [6], BrainSMASH [7], MENGA [8], Virtual Histology [9], or GCEA [10].
- **NiSpace integrates prior approaches into a unified framework**, allowing for combinations of, e.g., subject- and group-level analyses, uni- and multivariate methods, or autocorrelation-preserving group, map, and set permutations.
- The Python API is under development; a GUI will be available in the future.

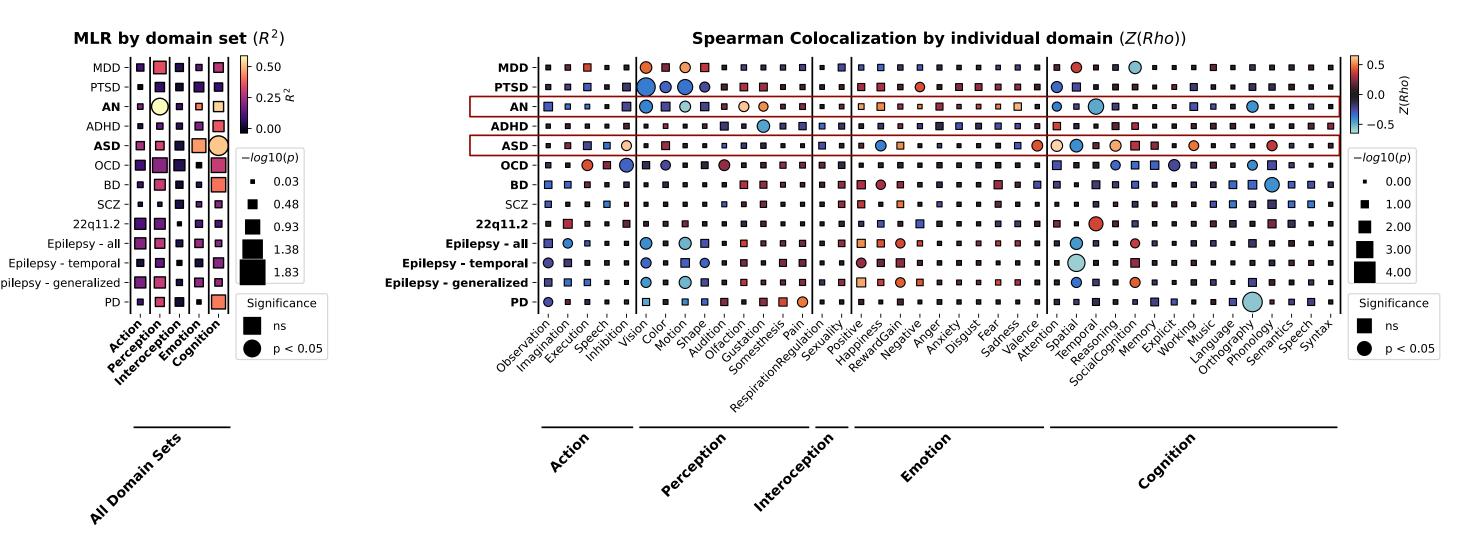


#### **ENIGMA** cortical thickness effect sizes – Map permutation

Colocalization between ENIGMA effect sizes maps and BrainMap behavioral domains in a multilevel analysis: MLR by domain class, followed by individual analyses for

each specific domain. AN (anorexia nervosa): perception; ASD: cognition.

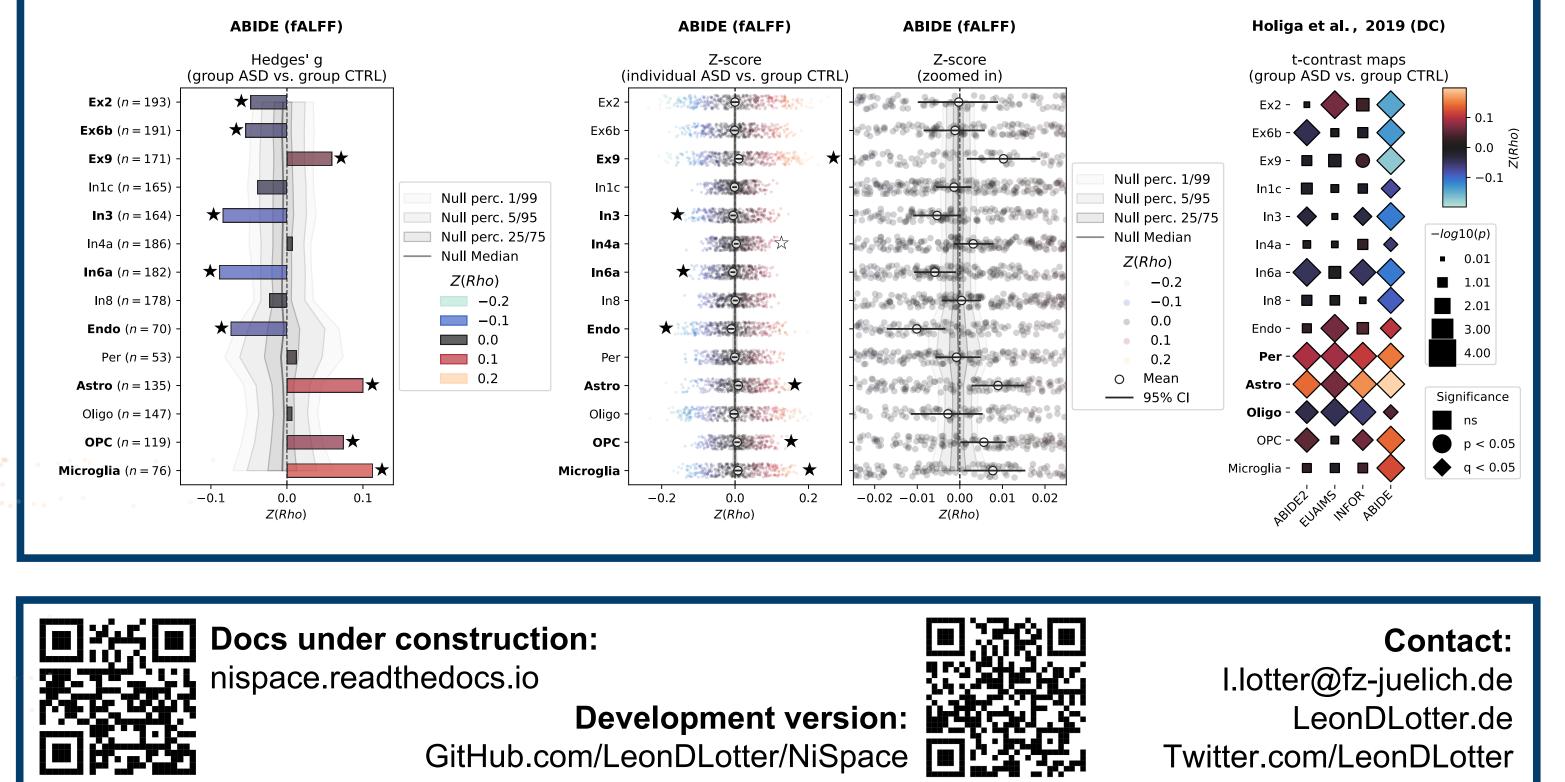
**Target**: ENIGMA effect size maps [mostly 5] | **Reference**: meta-analytic maps calculated from BrainMap coordinates [15] (included in **NiSpace**) | **Parcellation**: Desikan-Killiany | **Colocalization**: Multivariate regression (MLR); Spearman's Rho | Permutation: Spatial autocorrelation-preserving reference null maps ("Moran" method [4,6]).



#### Autism spectrum disorder (ASD) – Gene ("X")-set enrichment analysis

Colocalization between ASD-related resting-state brain activity/connectivity alterations and brain cell type markers: Consistent colocalization with astrocyte markers.

Target: ABIDE-I fALFF maps [12]; Group level DC t-contrast maps from four ASD cohorts [13] | Reference: Cell type marker genes [14] (included in **NiSpace**) | **Parcellation**: Schaefer-200+Melbourne-S2 | **Processing**: GM regression across parcels; age/sex regression & ComBat harmonization across subjects | Comparison: group level: parcel-wise Hedges' g (left); individual level: parcel-wise Z-scores (center) | Colocalization: Spearman's Rho (set-wise signed average) | Permutation: gene sets retaining original set sizes, sampled from the full (n ~= 7000) mRNA dataset.



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References: [1] Lotter, L. D. et al. (2024). In revision at Nat. Comm. [2] Dukart, J. et al. (2021). Nat. Neurosci. [4] Markello, R. D., Hansen J. Y. et al. (2022). Nat. Methods. [5] Larivière, S. et al. (2021). Nat. Methods. [6] Vos de Wael, R. et al. (2020). Comm. Biol. | [7] Burt, J. B. et al. (2020). NeuroImage. | [8] Rizzo, G. et al. (2018). Cereb. Cortex. | [10] Fulcher, D. B. et al. (2021). Nat. Comm. | [11] Dukart, J. et al. (2018). Sci. Rep. | [12] Craddock, C. et al. (2018). Cereb. Cortex. | [10] Fulcher, D. B. et al. (2021). Nat. Comm. | [11] Dukart, J. et al. (2018). Sci. Rep. | [12] Craddock, C. et al. (2018). Cereb. Cortex. | [10] Fulcher, D. B. et al. (2021). Nat. Comm. | [11] Dukart, J. et al. (2018). Sci. Rep. | [12] Craddock, C. et al. (2018). Cereb. Cortex. | [10] Fulcher, D. B. et al. (2021). Nat. Comm. | [11] Dukart, J. et al. (2018). Sci. Rep. | [12] Craddock, C. et al. (2018). (2013). Neuroinformatics 2013. [13] Holiga, S. et al. (2019). Sci. Transl. Med. [14] Lake, B. B. et al. (2018). Nat. Biotech. [15] Fox, P. T. et al. (2005). HBM.

Acknowledgments: Leon D. Lotter was supported by BMBF and MPG, Germany. | Original drug challenge study funded by F. Hoffmann-La Roche AG. | ABIDE-I data provided by the 1000 Functional Connectomes Project with funding from the NIMH.